# Digital Repository Universitas Jember Journal of **Biological** Research

Bollettino della Società Italiana di Biologia Sperimentale

www.jbiolres.org

#### **Editorial Board**

#### **Editors-in-Chief**

#### Marco Giammanco

Department of Surgical, Oncological and Oral Sciences, University of Palermo *Palermo, Italy* 

#### Gian Luigi Mariottini

University of Genova (retired), Research Fellow Department of Earth, Environment and Life Sciences *Genova, Italy* 

#### **Associate Editors**

Francesca Arfuso, Department of Veterinary Sciences, University of Messina, Messina, Italy Gaia Di Timoteo, Department of Biology and Biotechnology Charles Darwin, Sapienza University of Roma, Roma, Italy Alessia Fiorino, Department of Surgical, Oncological and Oral Sciences, Section of Medical Oncology, University of Palermo, Palermo, Italy Filippo Macaluso, Department of Biomedicine, Neuroscience and Advanced Diagnostics (BiND), Section of Human Anatomy, University of Palermo, Palermo, Italy Simona Manuguerra, Laboratory of Marine Biochemistry and Ecotoxicology, Department of Earth and Sea Sciences, University of Palermo, Palermo, Italy

Carmen Rizzo, National Institute of Biology, Department of Marine Biotechnology, Stazione Zoologica Anton Dohrn, Messina, Italy

#### Honorary Editors

Renzo Antolini, University of Trento (retired), Trento, Italy Massimo Cocchi, University of Bologna (retired), Bologna, Italy Luigi Pane, University of Genova (retired), Genova, Italy Emma Rabino Massa, University of Torino (retired), Torino, Italy

#### **Edit**orial Board

James Anthony, Michigan State University, East Lansing, USA Saeme Asgari, Pasteur Institute, Iran Han Bao, MSU-DOE Plant Research Laboratory of Michigan State University, USA Emilia Bellone, University of Genoa, Italy Maria Grazia Bridelli, University of Parma, Italy Dario Cantino, University of Turin, ItalyFrancesco Cappello, University of Palermo, ItalyDavid Caramelli, University of Florence, Italy Francesco Cappello, University of Palermo, Italy David Caramelli, University of Firenze, Italy Giuseppe Caramia, G. Salesi Hospital, Ancona, Italy Emilio Carbone, University of Turin, Italy Brunetto Chiarelli, University of Florence, Italy Pierluigi Consolo, University of Messina, Italy Amelia De Lucia, University "Aldo Moro", Bari, Italy Danila Di Majo, University of Palermo, Italy Luciano Fadiga, University of Ferrara, Italy

Caterina Faggio, University of Messina, Italy Vittorio Farina, University of Sassari, Italy Sara Ferrando, University of Genoa, Italy William Galanter, University of Illinois, Chicago, USA Lorenzo Gallus, University of Genoa, Italy Valerio Gennaro, ISDE Doctors for Environment, Genova, Italy Darren Grice, Institute for Glycomics and School of Medical Science, Griffith University, Nathan, Australia Stefania Grimaudo, University of Palermo, Italy Millie Hughes-Fulford, University of San Francisco, USA Gaetano Leto, University of Palermo, Italy Gianni Losano, University of Turin, Italy Mansoor A. Malik, Howard University Hospital, Washington DC, USA Herbert Ryan Marini, University of Messina, Italy Angela Marino, University of Messina, Italy Neville A. Marsh, Queensland University of Technology, Brisbane, Australia Bruno Masala, University of Sassari, Italy Alejandro M.S. Mayer, Midwestern University, Downers Grove, USA Concetta Maria Messina, Department of Earth and Sea Sciences, University of Palermo, Italy Vincenzo Mitolo, University "Aldo Moro", Bari, Italy Amir Sasan Mozaffari Nejad, Hamadan University of Medical Sciences, Iran Werner E.G. Muller, Johannes Gutenberg University, Mainz, Germany Giuseppe Murdaca, University of Genoa, Italy Giuseppe Palumbo, University Federico II, Naples, Italy Gian Luigi Panattoni, University of Turin, Italy Antonella Pantaleo, University of Sassari, Italy Massimo Pregnolato, University of Pavia, Italy Mark R. Rasenick, University of Illinois, Chicago, USA Angela Maria Rizzo, University of Milan, Italy Giacomo Rizzolatti, University of Parma, Italy Aldo Rustioni, University of North Carolina, USA Salvatore Sapienza, University of Catania, Italy Pietro Scotto Di Vettimo, University of Naples, Italy Vinicio Serino, University of Siena, Italy Lynne Christine Weaver, University of Western Ontario, Canada Ming Wei, Griffith University, Australia Mario Wiesendanger, University of Friburg, Switzerland

## Journal of Biological Research

is the official journal of the Società Italiana di Biologia Sperimentale and it is published online by <u>PAGEPress</u><sup>®</sup>, Pavia, Italy. All credits and honors to <u>PKP</u> for their <u>OJS</u>.

eISSN 2284-0230 pISSN 1826-8838



Published: 2021-01-01

**ORIGINAL ARTICLES** 



SCREENING OF HALOTOLERANT MICROFUNGI ISOLATED FROM HYPERSALINE SOILS OF ALGERIAN SAHARA FOR PRODUCTION OF HYDROLYTIC ENZYMES Wassila Dendouga, Mohamed Belhamra

RS

https://doi.org/10.4081/jbr.2022.10167

PDF



ANTIOXIDANT AND ANTIFUNGAL ACTIVITIES IN VITRO OF ESSENTIAL OILS AND EXTRACTS OF TWELVE ALGERIAN SPECIES OF THYMUS AGAINST SOME MYCOTOXIGENIC ASPERGILLUS GENERA Yamina Ben Miri, Aldjia Taoudiat, Mohamed Mahdid • https://doi.org/10.4081/jbr.2022.10299



WATER CONTENT, RESORPTION OF N AND P, AND THE GROWTH OF TEAK TECTONA GRANDIS L.F. SEEDLINGS ON FOUR TYPES OF GROWING MEDIA UNDER DROUGHT STRESS Slamet Santosa, Eddy Soekendarsi, Dody Priosambodo, Abdul Hayat Kasim

https://doi.org/10.4081/jbr.2022.9715



0

FIRST REPORT OF MICROALGAE RHEXHINEMA PAUCICELLULARE (ULVOPHYCEAE) IN MAURITIUS AND ITS BIOCHEMICAL EVALUATION AS A SOURCE OF FATTY ACIDS Ritesh Bhagea, Vishwakalyan Bhoyroo, Daneshwar Puchooa https://doi.org/10.4081/jbr.2022.9950 PDF

MOLECULAR DOCKING ANALYSIS OF SEAGRASS (ENHALUS ACOROIDES) PHYTOCHEMICAL COMPOUNDS AS AN ANTIDIABETIC Yayuk F<mark>atmaw</mark>ati, Sop<mark>hi San</mark>drina, Richca Nur Aina, Erlia Narulita https://doi.org/10.4081/jbr.2022.10224

## **REVIEW ARTICLES**



BIOACTIVE EFFECTS OF CITRUS FLAVONOIDS AND ROLE IN THE PREVENTION OF ATHEROSCLEROSIS AND CANCER Marco Giammanco, Fulvio Plescia, Manfredi M. Giammanco, Gaetano Leto, Carla Gentile https://doi.org/10.4081/jbr.2022.10313

BEY

Journal of Biological Research

ollettino della Società Italiana di Biologia Sperimentale

Molecular docking analysis of seagrass (Enhalus acoroides) phytochemical compounds

## as an antidiabetic

Yayuk Fatmawati,<sup>1</sup> Sophi Sandrina,<sup>1</sup> Richca Nur Aina,<sup>1</sup> Erlia Narulita<sup>1,2</sup>

<sup>1</sup>Department of Biology Education; <sup>2</sup>Center for Development of Advance Science and

Technology, University of Jember, Jember, East Java, Indonesia

Correspondence: Erlia Narulita, Center for Development of Advance Science and

Technology, University of Jember, Jember, East Java, 68121, Indonesia.

Tel.: +6282247777676

E-mail: erlia.fkip@unej.ac.id

Key words: Diabetes mellitus; molecular docking; phytochemicals; seagrass.

Acknowledgements: Authors would like to thank to Dr. Satya Ari Nugraha who assisting in molecular docking application.

**Contributions:** YF, SS, RNA and EN organize and design research, conduct research, provide research tools, collect and organize data. YF and RNA analyze and interpret data. SS and EN wrote the initial and final drafts of the article. All authors have critically reviewed

This article has been accepted for publication but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the final one. Please cite this article as doi: 10.4081/jbr.2022.10224

and approved the final draft and are responsible for the manuscript's content and similarity index. EN supervised the project.

**Fundings:** This research was funded by a grant from the Ministry of Education and Culture of the Republic of Indonesia with contract number of 1686/E2/TU/2020.

Conflict of interest: The authors declare no conflicts of interest.

## Abstract

*Enhalus acoroides* have potential to inhibit the  $\alpha$ -glucosidase enzyme and as an antidiabetic drug. Twenty-seven phytochemical compounds of seagrass (*E. acoroides*) were analyzed by molecular docking method. All possible candidatecompounds predicted ADME pharmacokinetic properties using the swissADME website. A molecular docking analysis was carried out using the PyRx 0.8 Autodock Vina software. Furthermore, the interaction analysis between molecules was carried out using PyMOL software and the Discovery Studio Visualizer BIOVIA. There were 17 of the27 compounds which had the best potency as oral antidiabeticdrug candidates. The validation results showed that all ligands had aroot mean score deviation(RMSD) value <2Å with the best value of 0.0. The binding affinity with the strongest bond value was -9.2 (kcal/mol) on the NAMPT bond with tannin, while the weakest value was 40.01 at 314y ( $\alpha$ -glucosidase) with 3-methyl. The 2h6d receptor can bind to all ligands, and the  $\alpha$ -glucosidase receptor can bind to two test ligands. The docking method used

in this study was valid, and the phytochemical compounds of seagrass have the potential to be an alternative to antidiabetic drugs.

Keywords: Diabetes mellitus; molecular docking; phytochemicals; seagrass.



### **INTRODUCTION**

Diabetes mellitus type 2 is a chronic condition and is one of the major causes of mortality worldwide. The disease is caused by high blood sugar levels because the insulin produced by the body isinsufficient to process sugar in the bodyor body resists insulin.<sup>1</sup>The epidemic of diabetes mellitus and its accompanying complications poses a major threat to global health problems. In 2015, 1 in 11 (around 415 million) adults aged 20-79 years had diabetes mellitus.<sup>2</sup>Based on data from the World Health Organization (WHO) in 2019, it was recorded that people living with diabetes reached 422 billion. This number is predicted to continue to increase to reach 642 million people with diabetes mellitus in 2040.<sup>2</sup>Some 90% of people with diabetes have type 2 diabetes as a result of unhealthy lifestyles. According to the International Diabetes Federation (IDF), Indonesia is one of the countries with the seventh most diabetes mellitus sufferers worldwide after China, India, the United States, Pakistan, Brazil, and Mexico.<sup>3</sup>

The high prevalence of type 2 diabetes mellitus sufferers in Indonesia can be treated from an early age by paying attention to some of the initial symptoms that appeared, which are related to the effects of high blood sugar levels.<sup>4</sup> The value of blood sugar levels in diabetes mellitus patients was> 126 mg / dL at fasting blood sugar levels, and at the time of the test was> 200 mg / dL. Early symptoms of type 2 diabetes mellitus can also be detected through a postprandial glucose level test, a blood glucose test done 2 hours after eating.<sup>5</sup>

The recognition of the initial symptoms that appeared can speed up treatment. Some medical treatments widely used for diabetes mellitus sufferers are pharmacological therapy using synthetic drugs and insulin injection. Those treatments have several drawbacks, such as dyspepsia symptoms, cell resistance, allergies due to the immune response for insulin injection, and the high cost. Another treatment method used is metformin, an oral antidiabetic

drug for initial treatment. The drawback of administering this drug is that it causes nausea and bloating in sufferers.<sup>6</sup>

The seagrass *(Enhalus acoroides)* is a natural ingredient that can be used as an antidiabetic.<sup>7</sup>Itcontains bioactive compounds such as flavonoids, alkaloids, phytochemical compounds, and antioxidants.<sup>8</sup> These compounds are known for reducing high blood sugar levels.<sup>9</sup> Delaying sugar absorption through inhibiting carbohydrate hydrolysis enzymes  $\alpha$ -glucosidase can control glucose levels in the body.<sup>10</sup>

The enzyme  $\alpha$ -glucosidase plays a role in the final step in the digestive carbohydrates,<sup>3</sup> and it worksto hydrolyze carbohydrates into sugars that are easier to absorb.<sup>11</sup>The inhibition of  $\alpha$ -glucosidase causes the enzyme cannot convert complex carbohydrates into simple sugarsand reduce the absorption in the small intestine. So, the result is to decrease postprandial plasma glucose levels and suppression postprandialhyperglycemia (PPHG).<sup>3</sup>Thus, it will reduce glucose levels in people with type 2 diabetes mellitus.<sup>12</sup>

## MATERIALS AND METHODS

#### **Receptor Structure Preparation**

The tools used in this study were PyRx 0.8 AutoDoc Vina software, PyMOL, and the Discovery Studio Visualizer 2020. The materials used in this study included 2D and 3D receptors and ligand structures. The research began by downloading the receptor structures, namely 3W37, 3A4A, and 3L4Yvia http://www.rcsb.org/pdb/home/home.do. These receptors are proteins with complex structures of the  $\alpha$ -glucosidase receptor. Then, three insulin receptors, namely 2H6D, 4WQ6, and 1BHS were chosen. The receptors were obtained by downloading the receptors' 3D structure on the RCSB PDB (The Research Collaboratory for Structural Bioinformatics Protein Data Bank) website in PDB format (http://www.rcsb.org/pdb/home/home.do).

## Preparation of 3-Dimensional Structures of Compounds as Test Ligands

The phytochemical compounds of seagrass (*E. acoroides*) were downloaded via the PubChem website (https://pubchem.ncbi.nlm.nih.gov/)and in 2D and 3D structures. Files were downloaded in SDF (Spatial Data File)format and converted to PDB format using the SMILES Translator Online website.<sup>13</sup>

#### **ADME Predictions**

Prediction of the pharmacokinetic properties of Absorption, Distribution, Metabolism, and Excretion(ADME)wasconducted by analyzing phytochemical compounds using the swissADME website (http://www.swissadme.ch/index.php). In general, ADME parameters were used to assess the work-range capability of phytochemical compounds after oral administration. Seagrass (*E. acoroides*) phytochemical compoundshave the potential to be the best candidates for the drug with a topological polar surface area (TPSA) value of <70  $Å^2$ (Angstrom = 10<sup>-10</sup> m).<sup>14</sup>

## Validation with PyRx 0.8 Autodock Vina

A total of 17 seagrass phytochemical compounds (*E. acoroides*) had TPSA values <70 (Å<sup>2</sup>) validated using PyRx 0.8 Autodock Vina. Then, the test ligand re-docked to the target protein. The center of the frame was placed in the ligand center and covered all residue of the binding site. The docking conformation results were aligned with the negative ligand conformation results of crystallographic measurements expressed in the root mean square deviation (RMSD) value. Validation was confirmed if the RMSD value of the re-docked and crystallographic ligands were less than 2Å.

## Molecular visualization using PyMOL and Discovery Studio

The validation results were used PyRx 0.8 Autodock Vina, which has an RMSD value <2 (Å), then visualized using PyMOL and Discovery Studio Visualizer BIOVIA 2019. Visualization of PyMOL was carried out to see the bond distance in three-dimensional space.

Meanwhile, the interactions between molecules in two-dimensional space was determined by Visualization Discovery Studio Visualizer BIOVIA 2019.

#### RESULTS

#### ADME Prediction (Absorption, Distribution, Metabolism, and Excretion)

The phytochemical compounds of seagrass, which were potential as drug candidates, were analyzed using SwissADME (Table 1). SwissADME was used to assess the rangeability of compounds on oral administration.<sup>12</sup> As much as 17 of 27 phytochemical compounds of seagrass *(E. acoroides)* had the potential as drug candidates with a TPSA value <70 (Å<sup>2</sup>) (Table 2).

#### **Docking Validation**

Docking validation aimed to establish the docking method's validity versus conformational 3D ligands to the target protein. Docking validation is expressed by RMSD. The docking validation results showed that all ligands had an RMSD value <2Å with the best value of 0.0 (Table 3). Thus, the docking method used in this study was valid. The binding affinity with the strongest bond value was -9.2 (kcal/mol) on NAMPT (Nicotinamide Phosphoribosyltransferase)bonds with tannin. While the weakest value was 40.01 at 314y ( $\alpha$ glucosidase) with 6-fluoro-2-[4-(2-fluorophenyl)phenyl]-3-methylquinoline-4-carboxylic acid (Table 4).

#### **Molecular Visualization**

Visualization of ligand binding interactions with the target receptor was carried out to determine the chemical bonds that occur and its stability (Figure 1). The application used to visualize the process is Discovery Studio Visualizer 2020. There are several ligands as candidates and then visualized to obtain several ligands with stronger and more stable bond types (Figure 2 and 3). The figure in the form of a dashed line indicates an interaction or bond that occurs. These binding interactions can be in electrostatic interactions, hydrophobic

interactions, van der Waals interactions, halogens, and hydrogen bonds. Hydrogen bonding optimizes hydrophobic interactions at the ligand and receptor surfaces. This will increase the binding affinity of the complex molecule. Weak hydrogen bonds will make it easier to break the interaction between the ligand and the receptor, so that it can be exchanged with other ligands. Hydrophobic interactions play an important role in increasing the binding affinity between the ligand surface and the receptor. Binding affinity and drug efficacy correlate with hydrophobic interactions, which are enhanced through interactions on hydrogen.<sup>15</sup>Electrostatic interactions between proteins and ligands play an important role in optimal affinity and selectivity.<sup>16</sup>The interaction between protein and stable ligand can activate the  $\alpha$ -glucosidase enzyme to inhibit the absorption of complex carbohydrates.<sup>17</sup>

Identification between ligands and receptors is the key in drug designing to predict drug candidates regarding the potential and side effects of a drug, namely the active potential of a compound that shows a good interaction between protein (receptor) and ligand. In molecular docking, there are two important things, namely structural data as receptor and ligand candidates and the procedures used to model the bonds between receptors and ligands.<sup>13</sup>

TPSA <140 (Å<sup>2</sup>) represents good intestinal absorption. Meanwhile, the TPSA value <70 (Å<sup>2</sup>) indicates a good value for brain penetration. The ligands of seagrass that havepotential as good drug candidates were shown with TPSA values <70 (Å<sup>2</sup>), analyzed by six receptors, namely AMP-activated protein kinase, NAMPT, 11- $\beta$ -hydroxysteroid dehydrogenase 1, and three  $\alpha$ -glucosidase receptors to find out the RMSD value and binding affinity.<sup>13</sup> These receptors are fundamental proteins in type 2 diabetes mellitus. AMP-activated protein kinase is involved in the stimulation of glucose transport and fatty acid oxidation. NAMPT is an intracellular regulator of nicotinamide adenine dinucleotide (NAD), regulating the activity of

NAD-dependent enzymes.<sup>18</sup> NAMPT is able to modulate the processes involved in insulin resistance. 11  $\beta$ -hydroxysteroid dehydrogenase 1 causes insulin resistance through conversion of cortisone to cortisol.<sup>19</sup>  $\alpha$ -glucosidase is an enzyme present in the small intestine that catalyzes the breaking of  $\alpha$ -1,4-glycosidic polysaccharide (or disaccharide) bonds by concurrent conversion to glucose.<sup>20</sup>

The results of the analysis using Discovery Studio Visualizer 2020shows that there are 11 compounds with the hydrogen bond type with van der Waals forces.<sup>21</sup> The hydrogen bond is a type of bond that plays an important role in the biological activity of a compound. The characteristics of the constituent protons in the hydrogen bond are dynamic.<sup>22</sup> This shows that the type of bond and the conditions of interaction between the ligand and the receptor were stable.<sup>23</sup>

## **CONCLUSIONS**

Molecular studies show that phytochemical compounds of seagrass can inhibit  $\alpha$ glucosidase activity and have the potential to be antidiabetic drugs. Inhibitor  $\alpha$ -glucosidase are effective in lowering insulin release, insulin requirement, and some can lower plasma lipidsInhibition of the activity of  $\alpha$ -glucosidase will decrease blood sugar levels in the body. Further in vivo and in vitro tests and studies are needed to confirm the compound is responsible for this favourable effects and molecular mechanisms of seagrass phytochemical compounds as an antidiabetic drugs.

#### REFERENCES

- Akrom, Harjanti PD, Armansyah. Hypoglycemic effects of ethanol extract of sweet cassava (*Ipomoea batatas* P) (Eeukr) in swiss alloxan-induced mice. Pharm J2014;4:65-76.
- Zheng Y, Ley SH, Hu FB. Global aetiology and epidemiology of type 2 diabetes mellitus and its complications. Nat Rev Endocrinol 2018; 14: 88-98
- 3. Wresdiyati T, Sa'diah S, Winarto A, Febriyani V. Alpha-glucosidase inhibition and hypoglycemic activities of *Sweitenia mahagoni* seed extract. Hayati 2015;22:73-8.
- 4. Rina N, Antarsih. Efek etil asetat daun lamun (*Enhalus acoroides* (LF) Royle) terhadap kadar MDA dan GSH mencit jantan tua). J Pen Kes 2019;4:2-56.
- Bare Y, Maulidi A, Sari DRT, Tiring SSND. In silico study predicts the potential of
  6-gingerol as an inhibitor of c-jun n-terminal kinases (JNK). Mat SciNet J 2019;1:59 63.
- Colquitt RB, Colquhoun DA, Thiele RH. In silico modelling as physiologic system.
  Best Pract Res Cl An 2011;25:499-510.
- Amudha P, Vanitha V. Toxicological, biochemical and histopathological evaluation of the ethanolic extract of seagrass-*Enhalus acoroides* in albino wistar rats. Prog Biotechnol 2019;19:1-10.
- Rahakbauw ID, Watuguly T. Analysis of *Enhalus acoroides* seagrass leaves compound in the coastal waters of Waai Village, Central Maluku Regency. Biopendix2016;3:1-53.
- Ganesh J. Faunal associates, trace metal content and bioefficacy of seagrass *Enhalus* acoroides (LF) Royle, 1839 and *Halodule uninervis* (Forsk.) Asch. 1882 from Kattumavadi, Palk Bay, Tamil Nadu, India. Geo 2011.

- Yuniarto A, Selifiana N. Aktivitas inhibisi enzim alfa-glukosidase dari ekstrak rimpang bangle (*Zingiber cassumunar* Roxb.) secara in vitro. Med Pharm Ind2018;2:1-22.
- 11. Menajang FSI, Mahmudi M, Yanuhar U, Herawati EY. Evaluation of phytochemical and superoxide dismutase activities of *Enhalus acoroides* (Lf) Royle from the coastal waters of North Sulawesi, Indonesia. Vet World 2020;13:676-80.
- Zuhro F, Puspitasari E, Muslichah S, Hidayat MA. Inhibitor α-Glukosidase Ekstrak Etanol Daun Kenitu (Chrysophyllum cainito L.) (α-Glucosidase Inhibitor Activity of Ethanol Extract Kenitu Leaves (Chrysophyllum cainito L.)). Pus Kes 2016;4:1-7.
- 13. Wati W, Widodo GP, Herowati R. Prediction of pharmacokinetics parameter and molecular docking study of antidiabetic compounds from *Syzogium polyanthum* and *Syzygium cumini*. J Sci Appl Chem 2020;23:189-95.
- 14. Rajan K, Zielensy A, Steinbeck C. STOUT: SMILES to IUPAC names using neural machine translation. Chem J 2021;13:1-14.
- 15. Patil R, Das S, Stanley A, Yadav L. Optimized hydrophobic interactions and hydrogen bonding at the target-ligand interface leads the pathways of drug-designing. PLOS ONE 2010;5:12-5.
- Rathi P, Ludlow F, Verdonk M. Practical high-quality electrostatic potential surfaces for drug discovery using a graph-convolutional deep neural network. Med Chem J 2019;63:8778-90.
- Frimayanti N, Zamri A, Eryanti Y, et al. Docking and molecular dynamic simulations study to search curcumin analogue compounds as potential inhibitor gainst SARS-C0V-2: a computational approach. J Sci Appl Chem 2021;3:85-90.

- Rachmania RA. Validation of virtual screening protocols and analysis of naturalbased cancer cell antiproliferation inhibitor interactions against cyclin-dependent kinase 4 (CDK4) receptors. Med Pharm J 2019;16:21-40.
- Ibrahim MA, Bester MJ, Neitz AW, Gaspar ARM. Rational in silico design of novel α-glucosidase inhibitory peptides and in vitro evaluation of promising candidates. Biomed Pharmacoter J2018;107:234-42.
- 20. Medina, KD, YS Moreno, D. Milenkovic, et al. In silico analysis of antidiabetic potential of phenolic compounds from blue corn (*Zea mays* L.) and black bean (*Phaseolus vulgaris* L.). Heliyon J 2020;6:1-3.
- 21. Morais FS, Canuto KM, Riberio PR, et al. Chemical profiling of secondary metabolites from *Himatanthus drasticus* (Mart.) Plumel latex with inhibitory action against the enzymes α-amylase and α-glucosidase: In vitro and in silico assays. Ethnopharmacologie 2020;253:1-9.
- 22. Vinsiah R, Fadhillah. Study of hydrogen bonding in the methanol-methanol and ethanol-ethanol system using the dynamic molecular method. Mat Nat Science J 2018;15:1-14.
- 23. Ali M, Khan KM, Mahdavi M, et al. Synthesis, in vitro and in silico screening ff 2amino-4-aryl-6-(phenylthio) pyridine-3,5-dicarbonitriles as novel a-glucosidase inhibitors. Bioorg Chem2020;100:1-13.

## TABLES

## Table 1: Phytochemical compounds of seagrass (Enhalus acoroides)\*

Phytochemical Compounds	Reference			
7-oxo-1H-[1,2,4]triazolo[1,5-a]pyrimidine-6-carboxylic acid	Ganesh, 2011			
Ethyl 7-amino-[1,2,4]triazolo[1,5-a]pyrimidine-6-carboxylate	Ganesh, 2011			
Ethyl ester	Ganesh, 2011			
2,2,7,7-tetramethyltricyclo[6.2.1.0 <sup>1,6</sup> ]undec-5-en-4-one	Ganesh, 2011			
Benzenamine	Ganesh, 2011			
2,4,6-trimethyl-N-(2,4,6-trimethylphenyl)benzenesulfonamide	Ganesh, 2011			
Benzene	Ganesh, 2011			
1-isocyano-2-methyl-3-nitrobenzene	Ganesh, 2011			
Benzenesulfonic acid	Ganesh, 2011			
N-[(Z)- <mark>[(3Z)-3-hydro</mark> xyiminobutan-2-	Ganesh, 2011			
ylidene] <mark>amino]benze</mark> nesulfonamide				
Benzyl alcohol	Ganesh, 2011			
Dibutyl phthalate	Ganesh, 2011			
Fumaric acid	Ganesh, 2011			
2-O-(3,5-difluorophenyl) 1-O-undecyl oxalate	Ganesh, 2011			
Hydrazinecarbothioamide	Ganesh, 2011			
2-(phenylmethylene)	Ganesh, 2011			
[(2S)-2-[(2R)-4-hexadecanoyloxy-3-hydroxy-5-oxo-2H-furan-2-	Ganesh, 2011			
yl]-2-hydroxyethyl] hexadecanoate				
Methanone	Ganesh, 2011			
Phenol	Ganesh, 2011			
6-fluoro-2-[4-(2-fluorophenyl)phenyl]-3-methylquinoline-4-	Ganesh, 2011			

carboxylic acid	
Propanoic acid	Ganesh, 2011
Dimethyl (isopropyl) silyl ester	Ganesh, 2011
Trans-3-Ethoxy-b-methyl-b-nitrostyrene	Ganesh, 2011
Saponin	Rina and Antarsih,
	2017
Flavonoid	Amudhaet al., 2018
Quercetin	Menajanget al.,2020
Tannin	Amudhaet al., 2018

\*Table redacted using data from the papers indicated

JE

Compound	TPSA (Å <sup>2</sup> ).
Methanone	17.07
1,2,3,4,5,6-Hexahydro-1,1,5,5-Tetramethyl-2,4a-	12.53
Methanonaphthalen	
1-isocyano-2-methyl-3-nitrobenzene	0.00
2-phenylmethylene	38.66
3-5-difluorophenyl undecyl ester	52.60
6-fluoro-2-[4-(2-fluorophenyl)phenyl]-3-methylquinoline-4-	50.19
carboxylic acid	
Benzenamine	26.02
Benzene	0.00
Benzenesulfonic acid	62.75
Benzyl alcohol	20.23
Dibutyl phthalate	<mark>52.6</mark> 0
Ethyl ester	26.30
Methanone	17.07
Phenol	20.23
Propanoic acid	37.30
Trans-3-Ethoxy-b-methyl-b-nitrostyrene	55.05

## Table 2: TPSA values of the phytochemical compounds of seagrass (Enhalus acoroides)

## Tannin

## Table 3: RMSD and Binding Affinity values of seagrass phytochemical compounds

(Enhalus acoroides)			
Receptors	Ligand	RMSD	Binding
		(Å)	Affinity
			(kcal / mol)
2h6d (AMP-	Methanone	1,584	-7.4
activated			
protein <mark>kinase)</mark>			
	1,2,3,4,5,6-Hexahydro-1,1,5,5-	0.0	-6.9
	Tetramethyl-2,4a-Methanonaphthalen		
	1-isocyano-2-methyl-3-nitrobenzene	0.0	-3.6
	2-phenylmethylene	1,659	-7.2
	3-5-difluorophenyl undecyl ester	0.556	-6.8
	6-fluoro-2-[4-(2-fluorophenyl)phenyl]-3-	0.0	-4.2
	methylquinoline-4-carboxylic acid		
	Benzenamine	0.0	-4.2
	Benzene	0.038	-3.9
	Benzenesulfonic acid	0.0	-5.2

	Benzyl alcohol	0.0	-4.5
	Dibutyl phthalate	0.0	-6.4
	Ethyl ester	0.0	-3.7
	Phenol	0.0	-4.2
	Propanoic acid	0.0	-3.3
	Trans-3-Ethoxy-b-methyl-b-nitrostyrene	0.0	-6.8
	Tannin	0.029	-7.8
4wq6	Methanone	0.0	-7.0
(Nicotinamide			
phosph <mark>oribosy</mark>			
ltransfe <mark>rase</mark>			
(NAMP <mark>T))</mark>			
	1,2,3,4,5,6-Hexahydro-1,1,5,5-	0.0	-5.7
	Tetramethyl-2,4a-Methanonaphthalen		
	1-isocyano-2-methyl	0.0	-3.7
	2-phenylmethylene	0.0	-7.0
	3-5-difluorophenyl undecyl ester	0.977	-6.7
	6-fluoro-2-[4-(2-fluorophenyl)phenyl]-3-	0.0	-6.2
	methylquinoline-4-carboxylic acid		
	Benzenamine	1,799	-4.1

	Benzene	0.121	-3.8
	Benzenesulfonic acid	0.0	-5.0
	Benzyl alcohol	0.041	-4.6
	Dibutyl phthalate	0.1	-5.8
	Ethyl ester	0.0	-3.3
	Phenol	0.0	-4.1
	Propanoic acid	0.0	-3.2
	Trans-3-Ethoxy-b-methyl-b-nitrostyrene	0.0	-5.8
	Tannin	0.016	-9.2
1bhs (1 <mark>1 β-</mark>	Methanone	0.0	-6.7
hydroxy <mark>steroid</mark>			
dehydrog <mark>enase</mark>			
1)			
	1,2,3,4,5,6-Hexahydro-1,1,5,5-	0.0	-6.8
	Tetramethyl-2,4a-Methanonaphthalen		
	1-isocyano-2-methyl	0.0	-3.6
	2-phenylmethylene	0.0	-7.1
	3-5-difluorophenyl undecyl ester	0.068	-6.1
	6-fluoro-2-[4-(2-fluorophenyl)phenyl]-3-	0.0	-8.1
	methylquinoline-4-carboxylic acid		

	Benzenamine	0.0	-4.6
	Benzene	0.0	-3.8
	Benzenesulfonic acid	0.0	-5.6
	Benzyl alcohol	0.0	-5.0
	Dibutyl phthalate	0.0	-5.7
	Ethyl ester	0.0	-3.6
	Phenol	0.0	-4.7
	Propanoic acid	0.0	-3.7
	Trans-3-Ethoxy-b-methyl-b-nitrostyrene	0.0	-8.5
	Tannin	0.0	-6.3
3a4a (α-	Methanone	0.0	-7.3
glucosida <mark>se</mark> )			
	1,2,3,4,5,6-Hexahydro-1,1,5,5-	0.0	-7.4
	Tetramethyl-2,4a-Methanonaphthalen		
	3-nitrobenzene	0.0	-3.6
	2-phenylmethylene	0.0	-7.1
	3-5-difluorophenyl undecyl ester	0.0	-6.5
	6-fluoro-2-[4-(2-fluorophenyl)phenyl]-3-	0.0	-2.2
	methylquinoline-4-carboxylic acid		
	Benzenamine	0.023	-4.4

	Benzene	0.0	-3.9
	Benzenesulfonic acid	0.0	-5.6
	Benzyl alcohol	0.042	-4.7
	Dibutyl phthalate	0.0	-6.3
	Ethyl ester	1,046	-3.6
	Phenol	0.0	-4.6
	Propanoic acid	0.0	-3.4
	Trans-3-Ethoxy-b-methyl-b-nitrostyrene	0.0	-6.5
	Tannin	0.013	-8.2
3w37 ( <mark>α-</mark>	Methanone	0.0	-3.4
glucosid <mark>ase</mark> )			
	1,2,3,4,5,6-Hexahydro-1,1,5,5-	0.0	-3.7
	Tetramethyl-2,4a-Methanonaphthalen		
	1-isocyano-2-methyl-3-nitrobenzene	0.0	-3.1
	2-phenylmethylene	0.0	-2.6
	3-5-difluorophenyl undecyl ester	0.0	-3.7
	6-fluoro-2-[4-(2-fluorophenyl)phenyl]-3-	0.0	35.6
	methylquinoline-4-carboxylic acid		
	Benzenamine	0.0	-4.4
	Benzene	0.067	-3.9

	Benzenesulfonic acid	0.0	-4.1
	Benzyl alcohol	0.0	-3.6
	Dibutyl phthalate	0.0	-4.0
	Ethyl ester	0.0	-3.8
	Phenol	0.0	-4.7
	Propanoic acid	0.0	-4.2
	Trans-3-Ethoxy-b-methyl-b-nitrostyrene	0.0	-4.4
	Tannin	0.0	-3.9
314y (α-	Methanone	0.0	-3.7
glucosi <mark>dase</mark> )			
	1,2,3,4,5,6-Hexahydro-1,1,5,5-	0.0	-2.1
	Tetramethyl-2,4a-Methanonaphthalen		
	1-isocyano-2-methyl-3-nitrobenzene	0.0	-4.2
	2-phenylmethylene	0.0	-3.0
	3-5-difluorophenyl undecyl ester	0.0	1.2
	6-fluoro-2-[4-(2-fluorophenyl)phenyl]-3-	0.0	40.01
	methylquinoline-4-carboxylic acid		
	Benzenamine	0.01	-5.0
	Benzene	0.004	-4.5
	Benzenesulfonic acid	0.012	-6.3

Tanı	I EKS	0.002	0.1
Trar	ns-3-Ethoxy-b-methyl-b-nitrostyrene	0.0	-5.6
Prop	panoic acid	1,541	-3.9
Pher	nol	0.0	-5.1
Ethy	vl ester	0.0	-4.1
Dibu	utyl phthalate	0.0	-2.5
Ben	zyl alcohol	0.002	-5.5



Receptors	Ligands	Amino acid residue involved
2h6d	Methanone	Thr211, Cys209, Leu212, Tyr205,
		Phe214, Pro234, Pro213, Phe231,
		Ala206.
	2 phenylmethylene	Tyr232, Thr211, Cys209, Tyr205,
		Phe214, Ile233, Pro234, Leu237,
		Ala206, Leu245, Phe231, Wal230,
		Lys225, Asp215, Gly210, Leu207,
		Leu208
	Benzenesulfonic acid	Thr211, Asp16, Arg311, Ser275,
		Ile309, Tyr188, Ala379
	Ethyl ester	Tyr205, Phe214, Cys209, Ala206
4wq6	2 phenylmethylene	Asp219, Edo607, Ser241, Val242,
		Po4602, Arg196, Tyr18, Phe193,
		His191, Pro273, Pro307, Ala244,
		Ile <mark>351, Asp16, Arg311, Ser27</mark> 5, Ile309,
		Tyr188, Ala379.
	Tannin	Asp219, Edo607, Ser241, Val242,
		Po4602, Arg196, Tyr18, Phe193,
		His191, Pro273, Pro307, Ala244,
		Ile351

## Table 4: Interactions between ligands and target proteins

1bhs	Methanone	Leu111, Lys159, Leu162, Phe160,
		Cys156, Ser158.
	Benzenamine	Val115, Leu162, Ser158
3w37	Dibutyl phthalate	Ile821, Gly820.
	Tannin	Ile821, Gly820.
3a4a	Tannin	Arg213, Arg442, Glu277, His351,
		Asp352, Asp69, Asp215, Asp69,
		Туг72.
314y	1,2,3,4,5,6-Hexahydro-	Val398
	1,1,5,5-Tetramethyl-	
	2,4a-	
	Methanonaphthalen	
	Tannin	Val398

FIGURES

Figure 1



Figure 2



Figure 3



#### **Figure Caption**

**Figure 1:** (A) The ligand interaction model methanone with 2h6d; (B) receptors Interaction model of methyl 2-(benzylideneamino) benzoate ligands with 2h6d receptors;(C) The interaction model of the benzenesulfonic acid ligand with the 2h6d receptor; (D) The interaction model of the ethyl ester ligand with the 2h6d receptor

**Figure 2:** (A) The interaction model of the methyl 2-(benzylideneamino) benzoate ligands with the 4w6q receptor; (B) The interaction model of the tannin ligand with the 4w6q receptor; (C) The interaction model of the 1-isocyano-2-methyl-3-nitrobenzeneligand with the 1bhs receptor; (D) Model of the interaction of the benzenamine ligand with the 1bhs receptor **Figure 3:** (A) The interaction model of the dibutyl benzene-1,2-dicarboxylate ligand with the 3w37 receptor; (B) Model of the interaction of the tannin ligand with the 3w37 receptor; (C) The interaction model of the tannin ligand with the 3w37 receptor; (C) The interaction model of the tannin ligand with the 3w37 receptor; (C) The interaction model of the tannin ligand with the 3a4a receptor; (D) The interaction model of the 1,2,3,4,5,6 Hexahydro 1,1,5,5 Tetramethyl 2,4a-Methanonaphthalen ligand with the 3l4y receptor