

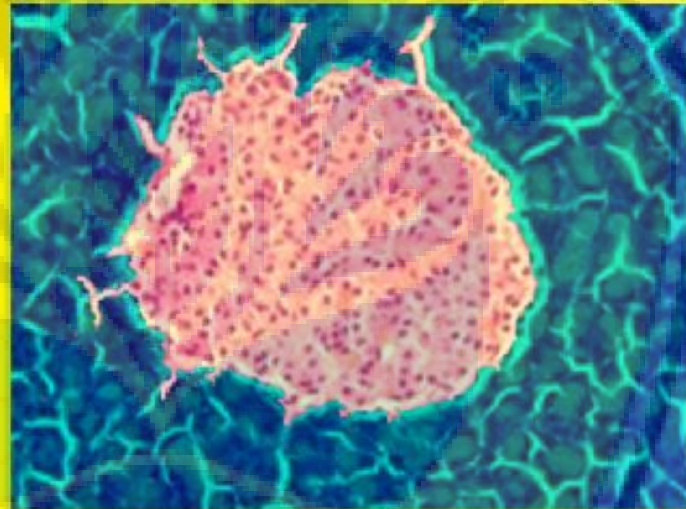
Indonesian. J. Pharm.
Volume 30 Issue 1 (2019)
January-March



ISSN : 2338-9427
Formerly ISSN : 0126-1037

Indonesian Journal of Pharmacy (*Indonesian J. Pharm.*)

Accredited by DGHE (DIKTI) No. 58/DIKTI/Kep/2013



STT NO. 1652/SK/DITJEN PPG/SST/1990

Faculty of Pharmacy
Universitas Gadjah Mada



Indonesian Journal of Pharmacy (ISSN-e: 2338-9486, ISSN-p: 2338-9427), formerly Majalah Farmasi Indonesia (ISSN: 0126-1037). The journal had been established in 1972, and online publication was begun in 2008. Since 2012, the journal has been published in English by Faculty of Pharmacy Universitas Gadjah Mada (UGM) Yogyakarta Indonesia in collaboration with IAI (Ikatan Apoteker Indonesia or Indonesian Pharmacist Association) and only receives manuscripts in English. Indonesian Journal of Pharmacy is Accredited by Directorate General of Higher Education (DGHE) DIKTI No. 58/DIKTI/Kep/2013.

Focus and Scope

The journal includes various fields of pharmaceuticals sciences such as:

- Pharmacology and Toxicology
- Pharmacokinetics
- Community and Clinical Pharmacy
- Pharmaceutical Chemistry
- Pharmaceutical Biology
- Pharmaceutics
- Pharmaceutical Technology
- Biopharmaceutics
- Pharmaceutical Microbiology and Biotechnology
- Alternative medicines

Journal Contact
Mailing Address

mfi@ugm.ac.id
Principal Contact

Prof. Dr. Sugiyanto, SU., Apt.
Prof. Dr.
Dept. of Pharmaceutical Chemistry, Faculty of Pharmacy, Gadjah Mada University,
Yogyakarta 55281, Indonesia
Phone: 0274-543120
Fax: 0274-543120
Email: mfi.ugm@gmail.com
Support Contact

Puma Arfah
Phone: 0274-543120
Email: mfi@ugm.ac.id

Editorial Team

Editor in Chief

1. [Prof. Sugiyanto Sugiyanto](#), Universitas Gadjah Mada, Department of Pharmacology and Clinical Pharmacy, Indonesia

Editorial Board

1. [Prof. Dr. Abdul Rohman](#), Department of Pharmaceutical Chemistry, Faculty of Pharmacy Universitas Gadjah Mada, Indonesia
2. [Prof. Dr. Shufeng Zhou](#), Department of Pharmaceutical Sciences, University of South Florida Tampa, United States
3. [Prof. Dr. Kazutaka Maeyama](#), Ehime University, Department of Pharmacology, Japan
4. [Prof. Dr. Masashi Kawaichi](#), Nara Institute of Science and Technology, Division of Gene Function in Animals, Japan
5. [Prof. Dr. Gunawan Indrayanto](#), Universitas Airlangga, Faculty of Pharmacy, Indonesia
6. [Prof. Dr. Veeresh P. Veerapur](#), Sree Siddaganga College of Pharmacy, Pharmaceutical Chemistry Department, India
7. [Prof. Dr. Agung Endro Nugroho](#), Universitas Gadjah Mada, Faculty of Pharmacy, Department of Pharmacology and Clinical Pharmacy, Indonesia
8. [Prof. Dr. Lee E. Kirsch](#), University of Iowa, Division of Pharmaceutics and Translational Therapeutics, United States
9. [Prof. Dr. Henk Timmerman](#), Vrije Universiteit Amsterdam, Division of Medicinal Chemistry, Netherlands
10. [Prof. Dr. Jeroen Kool](#), Vrije Universiteit Amsterdam, Division of BioAnalytical Chemistry, Netherlands
11. [Dr. Saikat Kumar Basu](#), University of Lethbridge, Department of Biological Sciences, Canada
12. [Dr. Joseph David Francis Tucci](#), La Trobe University, School of Pharmacy and Applied Science, Australia
13. [Dr. Chuda Chittasupho](#), Srinakharinwirot University, Department of Pharmaceutical Technology, Thailand
14. [Dr. Rina Kuswahyuning](#), Universitas Gadjah Mada, Faculty of Pharmacy, Department of Pharmaceutics, Indonesia
15. [Dr. Supang Khonde](#), University of Phayao, School of Pharmaceutical Sciences, Thailand
16. [Dr. Pudjono Pudjono](#), Universitas Gadjah Mada, Faculty of Pharmacy, Department of Pharmacology and Clinical Pharmacy, Indonesia
17. [Dr. Montarat Thavorncharoensap](#), Faculty of Pharmacy, Department of Pharmacy, Mahidol University, Thailand
18. [Dr. Karuna Shanker](#), Central Institute of Medicinal and Aromatic Plants India, Department of Analytical Chemistry, India
19. [Dr. Jun An](#), Sun Yat-Sen University, Department of Cardiothoracic Surgery, China
20. [Dr. Mohammed Emamussalehin Choudhury](#), Department of Pharmacology, Bangladesh Agriculture University, Bangladesh
21. [Dr. Abdul Wahab](#), Department of Pharmacy, Kohat University of Science and Technology (KUST), Pakistan

22. [Dr. Tony Hadibarata](#), Curtin University Sarawak Malaysia, Department of Environmental Engineering, Malaysia
23. [Dr. Shahin Gavanji](#), Department of Biotechnology, Faculty of Advanced Sciences and Technologies, University of Isfahan, Isfahan, Iran, Islamic Republic of



Reviewer

[Dr. Enade Perdana Istyastono](#), Faculty of Pharmacy, Universitas Sanata Dharma, Indonesia

[Dr. Susi Ari Kristina](#), Faculty of Pharmacy, Universitas Gadjah Mada, Indonesia

[Dr. Taha Nazir](#), Intellectual Consortium of Drug Discovery & Technology Development Inc., 937 Northumberland Ave Saskatoon Saskatchewan S7L3W8 Canada., Canada

[Dr. Gunawan Pamudji Widodo](#), Faculty of Pharmacy Setia Budi University Surakarta Indonesia, Indonesia

[Dr. Agatha Budi Susiana](#), Faculty of Pharmacy, Universitas Sanata Dharma, Indonesia

[Dr. Endang Lukitaningsih](#), Faculty of Pharmacy, Universitas Gadjah Mada, Indonesia

[Dr. Adam Hermawan](#), Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Universitas Gadjah Mada, Indonesia

[Dr. Uttam Budhathoki](#), Department of Pharmacy, Kathmandu University, Nepal

[Prof. Dr. Ridwan Amirudin](#), Faculty of Public Health, Universitas Hasanuddin, Indonesia

[Dr. Ari Sudarmanto](#), Faculty of Pharmacy, Universitas Gadjah Mada, Indonesia

[Dr. Dyah Aryani Perwitasari](#), Faculty of Pharmacy, Universitas Ahmad Dahlan., Indonesia

[Dr. Arief Nurrochmad](#), Faculty of Pharmacy, Universitas Gadjah Mada, Indonesia

[Prof. Dr. Shufeng Zhou](#), Department of Pharmaceutical Sciences, University of South Florida Tampa, United States

[Dr. Triana Hadna](#), Faculty of Pharmacy, Universitas Gadjah Mada, Indonesia

[Dr. Abdul Wahab](#), Department of Pharmacy, Kohat University of Science and Technology (KUST), Pakistan

[Dr. Montarat Thavorncharoensap](#), Faculty of Pharmacy, Department of Pharmacy, Mahidol University, Thailand

[Dr. Mohammed Emamussalehin Choudhury](#), Department of Pharmacology, Bangladesh Agriculture University, Bangladesh

[Dr. Dipak D Gadade](#), Shri Bhagwan College of Pharmacy, CIDCO N6, Aurangabad, India

[Dr I Wayan Mudianta](#), Ganesha University of Education, Bali, Indonesia, Indonesia

Digital Repository Universitas Jember

[Dr. Muthi Ikawati](#), Faculty of Pharmacy, Universitas Gadjah Mada, Indonesia

[Dr. Nining Sugihartini](#), Faculty of Pharmacy, Universitas Ahmad Dahlan, Indonesia



***In Vitro* Study of the Combination of Doxorubicin, *Curcuma xanthorrhiza*, *Brucea javanica*, and *Ficus septica* as a Potential Novel Therapy for Metastatic Breast Cancer**

Ika Rahmawati Sutejo^{1,3}, Herwandhani Putri³, Sri Handayani^{3,4}, Riris Istighfari Jenie^{2,3}, Edy Meiyanto^{2,3*}

1. Department of Biochemistry, Faculty of Medicine, University of Jember, Jalan Kalimantan 37 Jember 68121, Indonesia
2. Department of Pharmaceutical Chemistry Faculty of Pharmacy, Universitas Gadjah Mada, Sekip Utara Yogyakarta 55281, Indonesia
3. Cancer Chemoprevention Research Center, Faculty of Pharmacy, Universitas Gadjah Mada, Sekip Utara Yogyakarta 55281, Indonesia
4. Research Unit for Natural Product Technology, Indonesian Institute of Sciences (LIPI), Jalan Jogja - Wonosari, km 31.5, Kec. Playen, 174 WNO, Gading II, Gading, Playen, Kabupaten Gunung Kidul, Daerah Istimewa Yogyakarta 55861, Indonesia

Info Article

Submitted: 30-10-2018
Revised: 17-1-2019
Accepted: 13-3-2019

*Corresponding author
Edy Meiyanto

Author Correspondent
meiyan_e@ugm.ac.id

ABSTRACT

Less optimized therapeutic effects constrain the use of doxorubicin as the main agent of chemotherapy for metastatic breast cancer, resistance and side effects. Therefore we need a combination of more than one chemopreventive agent which has different molecular targets to solve that problem. The aims of this study is to prove the inhibitory effect of ethanolic extract of rhizome of *Curcuma xanthorrhiza* (ECx), fruit of *Brucea javanica* (EBj), leave of *Ficus septica* (EFs) and doxorubicin (Dox) alone and its combination on migration and invasion of a highly metastatic 4T1 breast cancer cell line. Cytotoxic activity of single and combination treatment was evaluated by MTT assay, followed by an experiment of apoptosis induction by using flow cytometry. The inhibitory effect on migration was observed by the scratch wound-healing assay. Furthermore, the observation of the activity of matrix metalloproteinase-9 (MMP-9) was analyzed by gelatin zymography. The results showed that ECx, EBj, EFs, and Dox has cytotoxic activity on 4T1 cells with the value of IC₅₀ respectively 49.7±1.53 µg/mL, 59.9±1.79 µg/mL, 15.2±2.12 µg/mL and 1.2±0.23 µM. Furthermore, the combination of ECx-EBj-Dox and ECx-EBj-EFs revealed a synergistic effect on 4T1 cells and decrease cell viability through the induction of apoptosis and necrosis. Based on wound healing assay, 24 hours incubation of this combination inhibited 4T1 cells migration compared to single treatment. Gelatin zymography analysis showed that this combination also inhibited the activity of MMP-9 greater than a single use. *Curcuma xanthorrhiza*, *Brucea javanica*, and *Ficus septica* may have potential to be developed as a combination with or without doxorubicin for metastatic breast cancer treatment.

Keywords: *C xanthorrhiza*, *B javanica*, *F septica*, antimetastasis, 4T1 cells.

INTRODUCTION

Breast cancer is the first leading cause of cancer deaths in women. Doxorubicin is an important modality for the treatment of breast cancer metastasis in addition to surgery (Bapsy and Sahoo, 2006). Long term use of doxorubicin

causes several side effects, resistance and toxicity to normal tissues (Smith *et al.*, 2010). Therefore, combining doxorubicin with the chemopreventive agent is needed to increase the activity of doxorubicin, to overcome the drug resistance and to reduce its side effect (Sarkar and Li, 2006).

The chemopreventive agent used in this research is a combination of *Brucea javanica* (EBj), *Ficus septica* (EFs) and *Curcuma xanthorrhiza* (ECx). Many clinical studies have suggested that *Brucea javanica* can be used alone as a conventional treatment for various cancers. However, the present study shows that *Brucea javanica* has synergistic effects when combined with certain anticancer drugs or radiotherapy (Nie *et al.*, 2012). The leaves of *Ficus septica* have been used to treat various cancers. The ethanolic extract of *Ficus septica* showed a cytotoxic effect on breast cancer T47D cell lines with an IC₅₀ value of 13 µg/mL. The extract at 4.88 µg/mL also showed an optimum synergistic effect in combination with doxorubicin (3.75 nM) (Pratama *et al.*, 2010). Besides, the extract induced apoptosis and downregulated the expression of Bcl-2 protein in breast cancer cells MCF-7 (Seki *et al.*, 2010). The principal components of *Curcuma xanthorrhiza* are curcumin and xanthorrhizol. Curcumin suppresses many key elements in cellular signal transduction pathways pertinent to growth, differentiation, and malignant transformation (Kunumakkara *et al.*, 2008). Choi *et al.* (2005) observed that injection of 0.2-1.0 mg/kg BW xanthorrhizol had an antimetastatic effect in a mouse lung metastasis model.

There have been several reports about the combination of doxorubicin with one kind of chemopreventive agent, and its synergistic has been proved (Lewandowska *et al.*, 2014), but study using a combination of more than one kind of chemopreventive agents with doxorubicin has not been done, mainly to prove its effectiveness as an antimetastasis agent in advanced breast cancer. This research combined ECx, and EBj with Dox to increase effectivity, to overcome drug resistance, and to reduce its side effect while the combination of three chemopreventive agents (ECx, EFs and EBj) without chemotherapy Dox is assumed to get nontoxic nature of EFs and to eliminate the side effect of Dox.

MATERIAL AND METHODS

The rhizome of *Curcuma xanthorrhiza*, the fruit of *Brucea javanica* and the leave of *Ficus septica* was macerated with 70% ethanol. The procedure was done at Medicinal Plant and Traditional Medicine, Research and Development Centre (B2P2TO-OT) Tawangmangu, Indonesia. Doxorubicin was obtained from Sigma. A DMSO (Merck) solution was used to dilute ethanolic

extract of ECx, EFs and EBj. The final DMSO concentration was set at less than 1%.

Cells culture

4T1 murine mammary carcinoma cells were acquired from Prof. Masashi Kawaichi (Nara Institute of Science and Technology, Japan) and maintained in Dulbecco's Modified Eagle's Medium (DMEM) high glucose containing Fetal Bovine Serum (FBS) 10% (v/v) (Sigma), penicillin-streptomycin 1% (v/v) (Gibco) and Fungizone 0.5% v/v (Gibco) in a humidified atmosphere of 5% CO₂ in air 37 °C.

Cytotoxicity assay

Various concentrations of samples (ECx, EBj, EFs, Dox, and its combination), DMEM culture medium (Gibco) with 10% FBS, 1% penicillin-streptomycin and fungizone were used to conduct the cytotoxic colorimetric MTT assay on 4T1 breast cancer cells. It was used to determine the IC₅₀ value. Afterwards, a cytotoxicity assay was also conducted to determine the effect of the combination of various sample concentrations treatment (Mosmann, 1983).

Flow cytometric apoptosis assay

Cells were added with Annexin-V-FLUOS Staining Kit (Roche) consisting of 200 µL of binding buffer, 2 µL of Annexin V, 2 µL propidium iodide (PI) and incubated for 10 minutes in the dark room according to manufacturer's instruction, then transferred into a flow cytometry tube and analyzed by flow cytometer (BD FACS Calibur, BD Bioscience) (Engeland *et al.*, 1998).

Scratch wound healing assay

The 4T1 cells (7.5×10^4) were cultured in 24-well plate and incubated for 24h. After starvation, cells were scratched by using sterile yellow tip then treated with various concentrations of samples. The cells were documented at 0, 18, and 24h (Liang *et al.*, 2007). The results were analyzed by using ImageJ software and converted to percentage closure parameter.

Gelatin zymography

The SDS-PAGE 8% supplemented with 0.1% gelatin was used to determine the activity of MMP-9 in the culture medium. After electro-phoresis running, gels were washed and incubated at room temperature. The reaction buffer was added and incubated for 24h at 37 °C. Gels were stained by using Coomassie Brilliant Blue R-250 (Sigma) and

destained until clear bands with dark blue background appeared (Kupai, 2011). The results were documented and analyzed by using Image J software.

Statistical analysis

Oneway ANOVA followed by the least significant difference (LSD) were used to assess the statistical significance. A statistically significant difference was considered to be present at $p < 0.05$. The results of scratch wound healing assay and gelatine zymography were documented and analyzed using Image J software before ANOVA.

Combination cytotoxicity assay and synergicity

The effect of combination treatment on 4T1 cells was evaluated by calculating the Combination Index (CI) value using the formula. Interpretation of the data was based on the classification (Reynolds and Maurer, 2005).

RESULTS AND DISCUSSION

Effects of ECx, EBJ, EFs, Dox alone and its combination on cell viability of 4T1

A single treatment of ECx, EBJ, EFs, or Dox showed cytotoxic effect on 4T1 breast cancer cells with IC_{50} value respectively $49.7 \pm 1.53 \mu\text{g/mL}$, $59.9 \pm 1.79 \mu\text{g/mL}$, $15.2 \pm 2.12 \mu\text{g/mL}$ and $1.2 \pm 0.23 \mu\text{M}$ (Figure 1). The reduction of cell viability of combination of ECx-EBJ-EFs and ECx-EBJ-Dox is greater than the single treatment. The combination of ECx-EBJ-Dox using concentrations of $\frac{1}{2} IC_{50}$ showed synergistic effects with a CI value of 0.63 and reduced cell viability up to 65% (Sutejo, 2015). The combination of ECx-EBJ-EFs using concentrations of $\frac{1}{2} IC_{50}$ showed synergistic effects with a CI value of 0.83 and reduced cell viability up to 63% (Figure 2).

The combination of ECx-EBJ-EFs and ECx-EBJ-Dox increase apoptosis and necrosis on 4T1 cells

The observation of apoptosis was done by using concentration $\frac{1}{3} IC_{50}$ of each agent. All single treatments and combination induced apoptosis on 4T1. The percentage of living cells in control was 92.18%, while the percentage of apoptotic cells is 6.34% and necrotic cells were 2.17%. After a single treatment ECx, EBJ, EFs, and Dox, the percentages of cells undergoing apoptosis were respectively 12.08%, 6.22%, 10.47% and 13.41%. The treatment with ECx-EBJ-EFs and ECx-EBJ-Dox for 24h incubation increased the apoptotic cells by 14.46% and 11.99% (Figure 3).

The combination of ECx-EBJ-EFs and ECx-EBJ-Dox inhibit migration of 4T1 cells

The observation of cell migration activity was done by using concentration $\frac{1}{4} IC_{50}$ of each agent. The effect on cell migration was observed at time point 0, 18, and 24h. The single treatment of ECx, EBJ, EFs, Dox and its combination decrease cell migration of 4T1. The ability of the most robust inhibition to the lowest showed by the combination of ECx-EBJ-EFs, ECx-EBJ-Dox, and then ECx, EFs, followed by EBJ, and the least one was doxorubicin (Figure 4).

The combination of ECx-EBJ-EFs and ECx-EBJ-Dox inhibit activity of MMP-9 on 4T1 cells

The treatment was done by using concentration $\frac{1}{4} IC_{50}$ of each agent. The single treatment of ECx, EFs, and EBJ is more potential to inhibit the activity of MMP-9 than doxorubicin. The combination of ECx-EBJ-EFs and ECx-EBJ-Dox resulted in much greater inhibition of MMP-9 activity than a single treatment (Figure 5).

Discussion

The single treatment of ECx, EBJ, EFs, and Dox was proven to have a cytotoxic effect on 4T1 cells (Figure 1), and its combination showed a synergism to reduce cell viability greater than a single treatment (Figure 2). It was supported by the previous study statements that each *Curcuma xanthorrhiza*, *Brucea javanica*, and *Ficus septica* increased the effects of chemotherapy (Kunnumakkara *et al.*, 2008). Combination of ECx-EBJ-EFs was as potent as ECx-EBJ-Dox to induce 4T1 cells death (Figure 3). Mechanism of cell growth inhibition both two combinations was due to apoptosis and necrosis pathways. The molecular mechanisms underlined the apoptosis after treatment of ECx, EBJ and EFs in 4T1 cells was through p53-independent pathways, due to 4T1 breast cancer cells line characteristic which is lack of protein P53 expression (Tao *et al.*, 2008). Whereas doxorubicin induces necrosis through increased $TNF\alpha$ and ROS (Sugimoto *et al.*, 2002).

The rhizomes of *Curcuma xanthorrhiza* contains volatile oil, saponin, flavonoid, and tannin. Chemistry analysis showed that the main substances of *Curcuma xanthorrhiza* are curcumin, demethoxycurcumin and xanthorrhizol (Choi *et al.*, 2005). Most of the studies on *Ficus septica* leaves reported on the phenanthroindolizidine

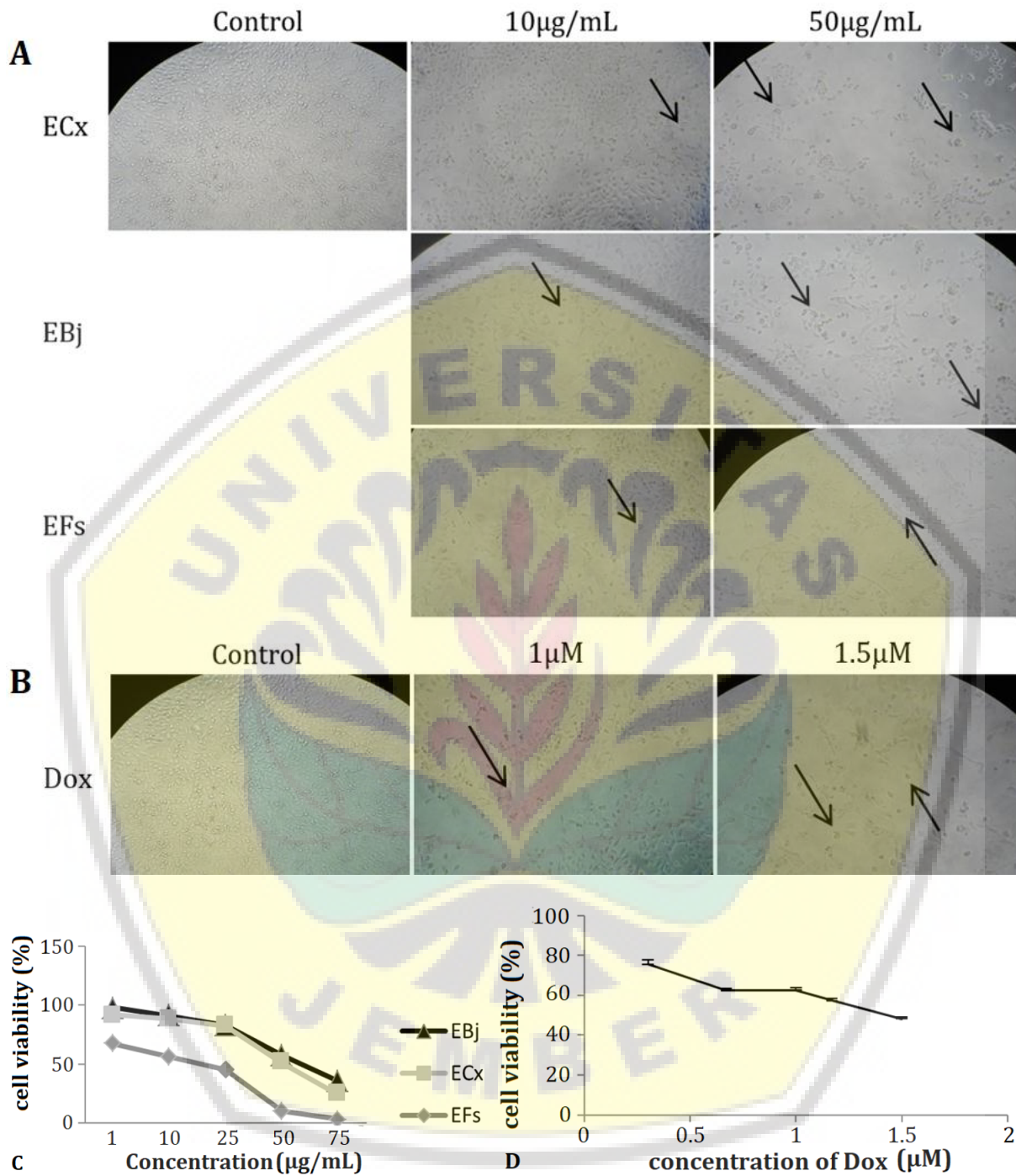


Figure 1. Cytotoxic effect of single treatment of ECx, EBj, EFs, and Dox on 4T1 Cells. 5×10^3 cells/well were seeded in 96 well plate and incubated for 24h, then treated with ECx, EBj, EFs and Dox. Cell viability was determined by using MTT assay as described in the method. Morphology cells after a single treatment of ECx, EBj, EFs (A) or Dox (B) for 24h. Arrows indicate cell morphological changes. Cell viability profile after a single treatment of ECx, EBj, EFs (C) or Dox (D) for 24h. Profile of cell viability were means \pm SD from 3 independent experiments.

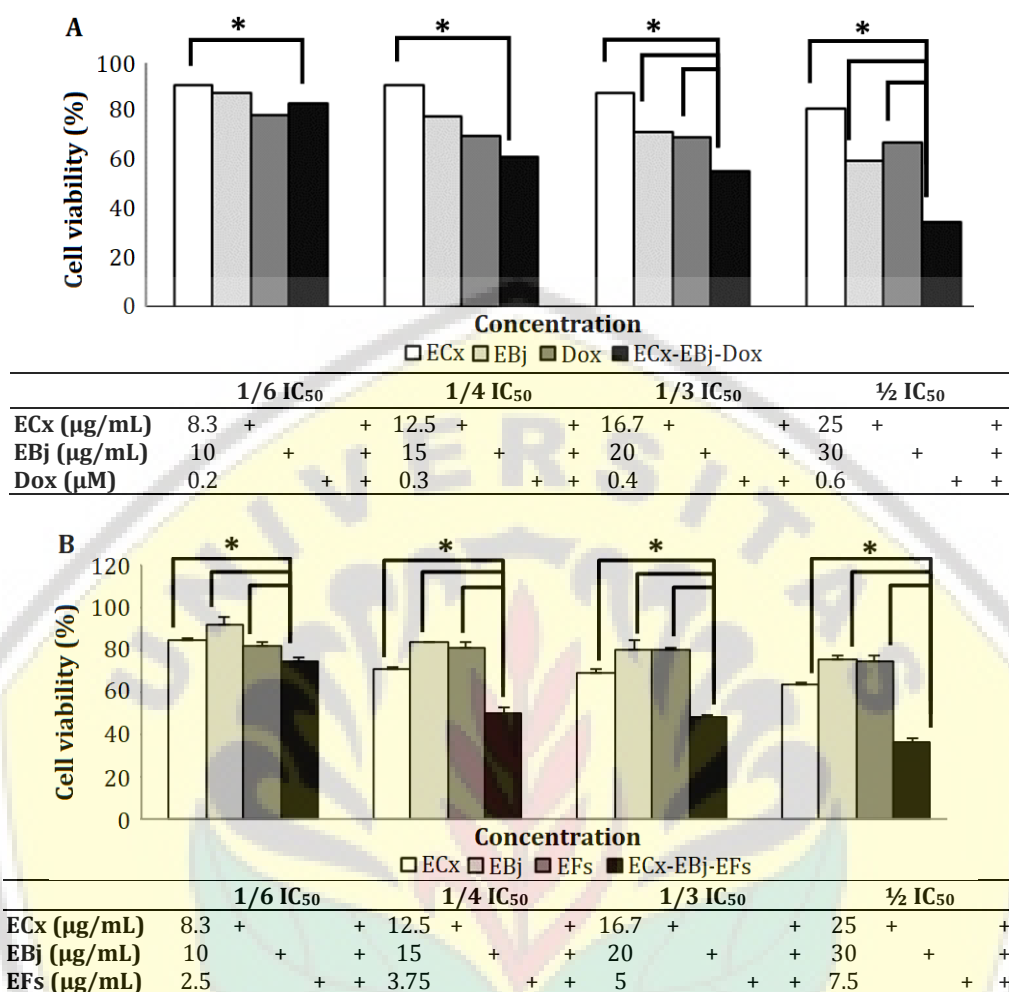


Figure 2. Combination effect of ECx, EBj, EFs, and Dox on the viability of 4T1 cells. 5×10^3 cells per well were incubated for 24h and exposed with various concentrations of (A) ECx, EBj, or Dox alone and its combination, (B) ECx, EBj, or EFs alone and its combination, then subjected for MTT assay. A combinational treatment yielded less cell viability compared to a single treatment ($p < 0.05$). Cytotoxicity was represented as a percentage of 4T1 cells viability as the mean \pm SE of three values.

alkaloid which has a cytotoxic effect on cancer cells (Seki *et al.*, 2010). Several natural components from *Brucea javanica* fruit include the tetracyclic triterpene quassinoids, anthraquinone, olein, oleic acid, linoleic acid, pregnane glucosides, and sesquiterpenes. In particular, tetracyclic triterpene quassinoids (brucein A and bruceantin) are the main active ingredients of *Brucea javanica* with remarkable antitumor activity (Chen *et al.*, 2013).

The treatment of ECx, EBj, EFs, and Dox alone or in combination inhibited the migration of 4T1 breast cancer cells (Figure 4). The interesting finding was that Dox as the main chemotherapy in advanced cancer has the lowest antimigration

activity compared to ECx, EBj and EFs (Figure 4). Bandyopadhyay *et al.* (2010) and Krstic and Santibanez (2014) proved that doxorubicin increases migration and invasion of 4T1 and MDAMB-231 cells through induction of TGF β . Those findings in line with our result that the combination of ECx-EBj-EFs inhibited cells migration greater than ECx-EBj-Dox. Metastasis inhibition of doxorubicin which was seen in this study was due to cell death caused by doxorubicin treatment. Although the mechanism is not clear, single doxorubicin proved to inhibit the M5076 ovarian cancer cell metastasis *in vivo* (Sugiyama and Sadzuka, 1999).

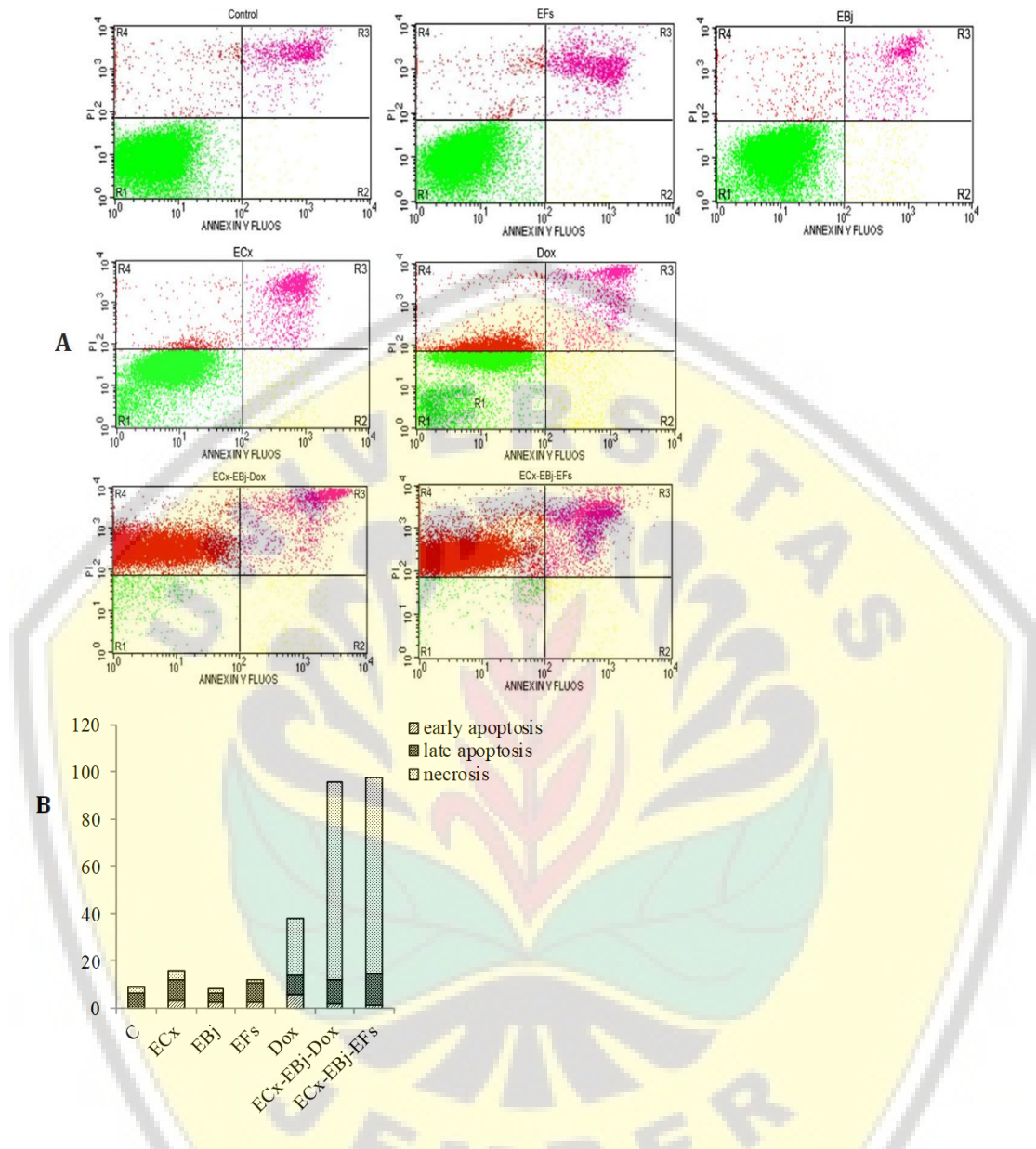


Figure 3. Apoptosis effects of ECx, EBJ, EFs, and Dox single and its combination on 4T1. Cells were harvested after treatment of 16.7 $\mu\text{g}/\text{mL}$ of ECx, 20 $\mu\text{g}/\text{mL}$ of EBJ, 5 $\mu\text{g}/\text{mL}$ EFs and 0.4 μM Dox for 24h single or in combination, then stained using Annexin V-FITC/PI and were analyzed by using flow cytometry. (A) The distribution profiles of living cells (R1), early apoptosis (R2), late apoptosis (R3), and necrosis (R4), (B) quantification of cells undergoing early apoptosis, late apoptosis and necrosis.

Several reports related to the anti-metastasis potency of the active compounds contained in the extracts we used are as follows. Curcumin as the main active ingredient of ECx, inhibit the migration of breast cancer cells MDAMB-231 by suppressing the FAK pathway and lowering the expression of PI3K and were subsequently able to decrease the expression of

VEGF (Lin *et al.*, 2009). Curcumin also inhibits the expression of MMP-9 causing a decrease of MMP-9 activity, inhibits β -catenin and reduce the loss of E-cadherin, which is related to the ability of invasion and metastasis of cancer cells (Thangapazham *et al.*, 2006). Alkaloids phenanthro-indolizidine from *Ficus septica* inhibits expression of COX-2 proteins (Mandhare *et al.*, 2015).

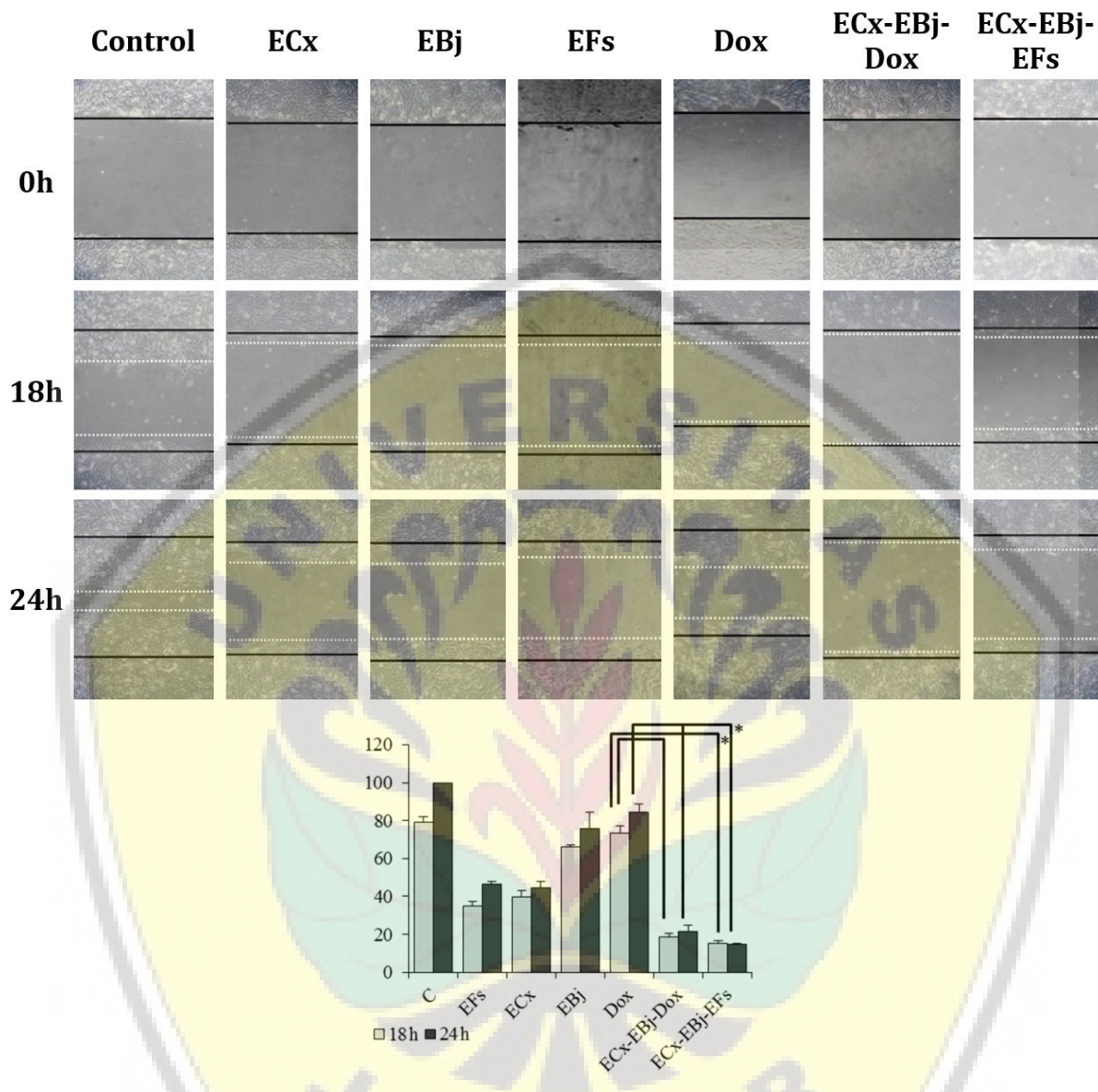


Figure 4. Effects of ECx, EBJ, EFs, and Dox alone and its combination on 4T1 cell migration. (A) The morphology of the cells after the scratch and treated with 12.5 $\mu\text{g}/\text{mL}$ ECx; 15 $\mu\text{g}/\text{mL}$ EBJ, 3.75 $\mu\text{g}/\text{mL}$ EFs, or 0.3 μM Dox single and its combination. The observation was made after 0, 18, 24, and 42h of treatment under an inverted microscope with 100x magnification. (B) The percentage of 4T1 closure after 18h and 24h of treatment. The value was a means of % closure \pm SD from 3 independent experiments. The area of the scratch was analyzed using ImageJ software then % closure was calculated following the procedure of the analysis. The asterisk (*) indicates differences ($p < 0.05$; $n = 3$)

Brucein A and bruceantin are known to inhibit the invasion and migration of tumour cells targeting at MRP-1/CD9 and integrin α -5 (Nan *et al.*, 2015). Brucein A from *Brucea javanica* lowered VEGF expression so that no signal is transmitted through the VEGF receptor (Xinjie and Linyi, 2013). Moreover, *Brucea javanica* and *Ficus septica* also

suppress the activation of NF- κ B and inhibits expression of COX-2 proteins that promote invasion (Kim *et al.*, 2010; Mandhare *et al.*, 2015). The mechanisms above explain our findings that the combination of ECx-EBJ-EFs and ECx-EBJ-Dox inhibited cells migration greater than the single treatment (Figure 4B).

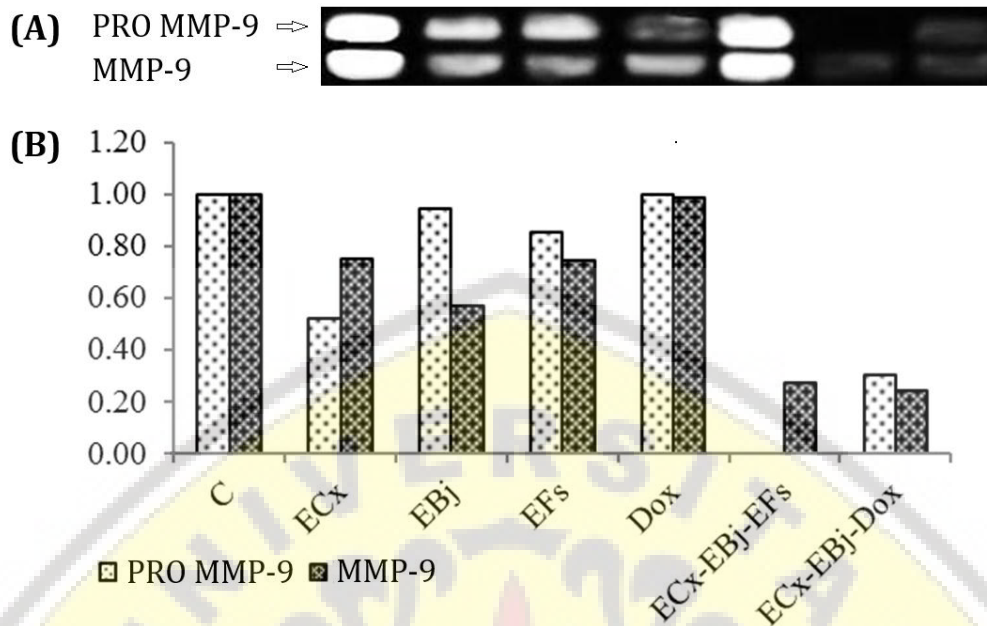


Figure 5. Effect of ECx, EBj, EFs, Dox alone & its combination in MMP-9 activity on 4T1 cells. Cells were treated with 12.5 $\mu\text{g}/\text{mL}$ ECx; 15 $\mu\text{g}/\text{mL}$ EBj, 3.75 $\mu\text{g}/\text{mL}$ of EFs, or 0.3 μM Dox and their combination. MMP-9 activity assay was conducted using gelatin zymography according to the method. Analysis of the results was done by using ImageJ software to measure the intensity of gelatin degradation by pro-MMP-9 & MMP-9 in the gel. (A) The bands that showed the pro MMP-9 & MMP-9 activities. (B) The graph of the relative intensity of pro-MMP-9 & MMP-9 compared to controls (n=3).

The fact that Dox has not only induced cell migration but also cell invasion also explains our data that the combination of ECx-EBj-EFs inhibited MMP-9 activity greater than ECx-EBj-Dox (Figure 5B).

CONCLUSION

The results of this study conclude that both two combinations of ECx-EBj-Dox and ECx-EBj-EFs have a cytotoxic effect via apoptosis and necrosis pathways. The combination of ECx-EBj-EFs has antimigration activity and inhibition of cancer cell invasion greater than ECx-EBj-Dox. Both two combinations have a potency to be developed as a co-chemotherapy agent in breast cancer metastasis. Nevertheless, further researches are still needed in order to apply such combination therapy for cancer patients.

ACKNOWLEDGEMENT

We gratefully thank The Ministry of Research, Technology, & Higher Education (Kemenristek-DIKTI) for funding this research through Insentif Riset Sistem Inovasi Nasional (Insinas) 2014 research grant.

REFERENCES

- Bandyopadhyay A., Wang L., Agyin J., Tang Y., Lin S., Yeh I., De K., and Sun L. 2010. Doxorubicin in combination with a small TGF β inhibitor: a potential novel therapy for metastatic breast cancer in mouse models. *Plos One*, 5(4):e10365.
- Bapsy PP., and Sahoo TP. 2004. Recent advances in the management of metastatic breast cancer, *Indian Journal of Medical & Paediatric Oncology*, 24 (2):19-26.
- Chen M., Chen R., Wang S., Tan W., Hu Y., Peng X., and Wang Y. 2013. Chemical components, pharmacological properties, and nanoparticulate delivery systems of *Brucea javanica*, *Int. J. nanomedicine*, 8. 85-92. 10.2147/IJN.S31636.
- Choi MA., Kim SH., Chung WY., Hwang JK., and Park KK. 2005. Xanthorrhizol, a natural sesquiterpenoid from *Curcuma xanthorrhiza*, has an anti-metastatic potential in experimental mouse lung metastasis model, *Biochem. Biophys. Res. Commun.* 326(1):210-7.

- Engeland M., Nieland LJW., Ramaekers FCS., Schutte B., and Reutelingsperger CPM. 1998. Annexin V-affinity assay: a review on an apoptosis detection system based on phosphatidylserine exposure, *Cytometry*, 31:1-9.
- Kim J., Lau EK., Pan L., and De Blanco EJC. 2010. NFκB inhibitors from *Brucea javanica* exhibiting intracellular effect on reactive oxygen species, *Anticancer res.*, 30: 3295-3300.
- Krstic J., and Santibanez JF. 2014. Transforming growth factor-beta and matrix metalloproteinases: functional interaction in tumor stroma-infiltrating myeloid cells, *The Scientific World Journal*, 2014: 521754.
- Kunnumakkara AB., Anand P., and Aggarwal B. 2008. Curcumin inhibits proliferation, invasion, angiogenesis & metastasis of different cancers through interaction with multiple cell signalling proteins, *Cancer Letter* 269: 199-225.
- Kupai K. 2011. Gelatin zymography for detection of matrix-metalloproteinase-2 and -9 (mmp-2, mmp-9) from myocardial samples, *Practical course: Basic biochemical methods and ischemic heart model* (link:http://www3.szote.u-szeged.hu/hurodocs/hun/downloads/biochemistry/HURO_zymo_practical_Kupai_final.pdf).
- Lewandowska U., Gorlach S., Owczarek K., Hrabec E., Szewczyk K. 2014. Synergistic interaction between anticancer chemotherapeutics and phenolic compounds and anticancer synergy between polyphenols, *Postepy Hig Med Dosw (online)*, (68):528-540.
- Liang C., Park AY., and Guan J. 2007. In vitro scratch assay: a convenient & inexpensive method for analysis of cell migration in vitro, *Nature Protocol*, 2 (2). Doi:10.1038/nprot.2007.30.
- Lin HJ., Su CC., Lu HF., Yang JS., Hsu SC., and Ip SW. 2010. Curcumin blocks migration & invasion of mouse-rat hybrid retina ganglion cells (n 18) through the inhibition of MMP-2, -9, PAK, Rho A & Rock-1 Gene expression, *Oncology Reports*, 23: 665-670 DOI: 10.3892/or_00000682.
- Mandhare AA., Dhulap SA., Dhulap AS., and Biradar SC. 2015. Review on the Anticancer and In-silico Binding Studies of Phenanthroindolizidine Alkaloids, *ImedPub journal*, Vol 1 No 1:5.
- Mosmann T. 1983. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays, *Journal of Immunological Methods*, 65: 55-63.
- Nan Z., Li YH., Wu XK., Wang GY., Cai DY., and Han FJ. 2015. Effect of *Brucea javanica* fruit oil emulsion combined cisplatin on the growth inhibition of transplanted tumor in human ovarian cancer SKOV3 nude mice: an experimental study. *Zhongguo Zhong Xi Yi Jie He Za Zhi*, 35 (1): 57-62.
- Nie YL., Liy KX., Mao XY., Li YL, Li J., and Zhang MM. 2012. Effect of injection of *Brucea javanica* oil emulsion plus chemoradiotherapy for lung cancer: a review of clinical evidence, *J Evid Based Med.*, 5(4):216-25.
- Pratama RH., Ikhtiarisyah YG., Fitriasari A., Anindyajati Ikawati M., Meiyanto E. 2011. Awar-awar leaves ethanolic extract synergistically enhances cytotoxic of doxorubicin on T47D breast cancer cells, *Ind J Pharm Sci.*, 9(1):67-71.
- Reynolds CP., and Maurer BJ. 2005. Evaluating response to antineoplastic drug combinations in tissue culture models, *Methods Mol Med.*, 110:173-83.
- Sarkar FH., and Li Y. 2006. Using Chemopreventive agents to enhance the efficacy of cancer therapy, *Cancer Rev.*, 66(7):3347-3350.
- Sekti DA., Mubarak MF., Armandani I., Junedy S., and Meiyanto E. 2010. Awar-awar (*Ficus septica* Burm. F.) leaves ethanolic extract induced apoptosis of MCF-7 cells by downregulation of Bcl-2, *J Trad Med.*, 15(3):100-104.
- Smith LA., Cornelius VR., Plummer CJ., Levitt G., Verill M., Canney P., and Jones A. 2010. Cardiotoxicity of anthracycline agents for the treatment of cancer: Systematic Review & Meta-analysis of Randomised. *BMC Cancer*, 10:337.
- Sugimoto K., Tamayose K., Sasaki M., Hayashi K., and Oshimi K. 2002. Low-dose doxorubicin-induced necrosis in Jurkat cells and its acceleration & conversion to apoptosis by antioxidants, *British Journal of Haematology*, 118:229-238. DOI: 10.1046/j.1365-2141.2002.03577.
- Sugiyama T., and Sadzuka Y. 1999. Combination of theanine with doxorubicin inhibits hepatic

- metastasis of M5076 ovarian sarcoma, *Clin. Cancer Res.*, 5, 413–416.
- Sutejo IR. 2015. Potensi Antimetastasis Kombinasi Ekstrak Etanolik Temulawak (*Curcuma xanthorrhiza*), Buah Makassar (*Brucea javanica*) dan Doxorubicin pada Sel 4T1, Thesis, Gadjah Mada University.
- Tao K., Fang M., Alroy J., and Sahagian GG. 2008. Imagable 4T1 model for the study of late-stage breast cancer, *BMC Cancer*, 8:228.
- Thangapazham RL., Sharma A., and Maheshwari RK. 2006. Multiple molecular targets in cancer chemoprevention by curcumin, *The AAPS Journal*, 8: E443.
- Xinjie DU., and Linyi HE. 2013. The study on the expression effects of Brucea Javanica oil on VEGF and bFGF in proliferative endometrium hyperplasia of rats, *Journal of Binzhou Medical University*, 2013-01.



