

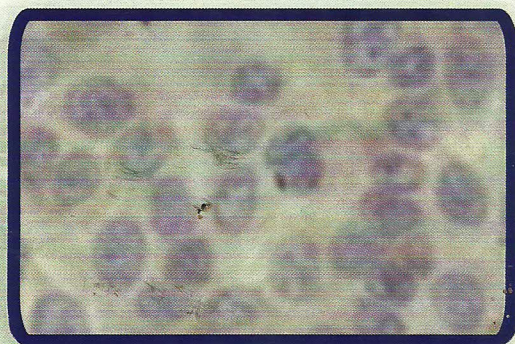
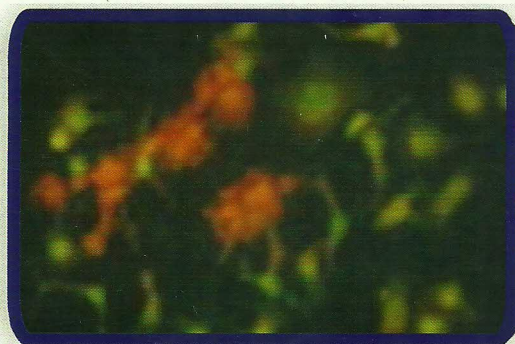


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(see Handayani et al. pages 318-324)



Combination of Leunca Herb Ethanolic Extract and Doxorubicin Suppresses HeLa Cells' Growth

Sarmoko^{1,2}, Dyaningtyas D. P. Putri², Endah Puspitasari^{2,3}, Anindyajati², and Edy Meiyanto^{2*}

¹Department of Pharmacy, Faculty of Medicine and Health Sciences, Universitas Jenderal Soedirman, Purwokerto, Jawa Tengah,

²Cancer Chemoprevention Research Center, Faculty of Pharmacy, Universitas Gadjah Mada, Yogyakarta,

³Faculty of Pharmacy, Universitas Jember, Jember, Jawa Timur.

Abstract

Leunca (*Solanum nigrum* L.) ethanolic extract showed cytotoxic activity on several cancer cell lines (HepG2, HT-29) and showed anti-proliferative activity on MCF-7 cells. Its application as a combination agent in chemotherapy will increase the effectivity and reduce the toxicity of chemotherapy. We predict that application of combinatorial chemotherapy in cancer treatment will be more effective and less toxic compared to single treatment. Our research aims to investigate the cytotoxic activity of leunca herbs ethanolic extract alone and in combination with doxorubicin on HeLa cell line. MTT assay was conducted to measure the growth inhibitory effect of leunca herbs ethanolic extract and combinatorial treatments. Leunca herb ethanolic extract (5, 50, 250 µg/ml) increased the cytotoxic effect of doxorubicin compared to doxorubicin alone. The strongest cytotoxic activity resulted from the combination of 250 µg/ml leunca herbs ethanolic extract and 250 nM doxorubicin. Based on our results, leunca herbs ethanolic extract is a potential chemopreventive agent, while its molecular mechanism needs to be explored.

Keyword : Leunca herbs ethanolic extract, doxorubicin, HeLa, MTT assay

INTRODUCTION

Combination chemotherapy (co-chemotherapy) has been used widely due to tumor cell heterogeneity, drug resistance, and the increasing successfulness in the clinic (Kufe *et al.*, 2003). Chemoprevention agents originated from nature seems to be a promising candidate for co-chemotherapeutic agents. Leunca (*Solanum nigrum* L.) is one of the herbs having this potency. It has solanine, solasodine, and solamargine (Everist, 1974; Weller and Phipps, 1979).

Solanine is able to induce apoptosis in HepG2 cells mediated by the inhibition of Bcl-2 expression (Ji *et al.*, 2008). While β-2-solamargine is cytotoxic on some cancer types, e.g.: colon cancer (HT-29 and HCT-15), prostate cancer (LNCaP and PC-3), and breast cancer (T47D and MDA-MB-231) (Hu *et al.*, 1999). Solamargine could also induce apoptosis through mitochondrial pathway (Liang *et al.*, 2008) and modulate the expressions of TNFRs and Bcl-2 on H441, H520, H661, and H69 human lung cancer

cells (Liu *et al.*, 2004). Thus, its potential as co-chemotherapeutic agent needs to be explored.

Leunca herb ethanolic extract has been found to have synergistic effect when combined with doxorubicin on T47D cells (Anindyajati *et al.*, 2010) as well as with cisplatin on HeLa cells (Istiaji *et al.*, 2010). This study aimed to observe the effect of leunca's ethanolic extract (LEE) application on the cytotoxicity performed by doxorubicin, analyzed by MTT assay.

The data could complete the co-chemotherapeutic potency of leunca. Combinatorial treatment of doxorubicin and LEE were applied in order to increase the cytotoxicity of doxorubicin on HeLa cells, allowing the use of lower dose of the chemotherapeutic agent giving less toxicity on normal tissues.

*Corresponding author email : meiyana_e@ugm.ac.id

METHODS

Sample preparation

Dried powder of leunca herbs were purchased from Balai Besar Penelitian dan Pengembangan Tanaman Obat dan Obat Tradisional (B2P2TOOT), Indonesia. Dried powder was then extracted by maceration for 5 days with 70% ethanol. Collected filtrate was concentrated using rotary evaporator (Heidolph WB2000), then dissolved in Dimethyl Sulfoxide (DMSO) (Sigma). Both 5mg/ml doxorubicin and extract solution were diluted in DMEM cell culture medium before being applied. DMSO was used as the co-solvent in dissolving LEE in DMEM culture medium.

Chemicals

Dulbecco's Modified Eagle Medium (DMEM) powder (Gibco), 10% Fetal Bovine Serum (FBS) (Gibco), and 10,000 units/ml Penicillin-10,000 µg/ml Streptomycin (Gibco) were used for cell culture medium. Cells were harvested using 25% Tripsin EDTA (Gibco). For cytotoxicity assay, 10% Sodium Dodecyl Sulphate (SDS) (Merck) dissolved in 0.1N HCl (Merck) as stopper reagent, and 3-[4,5-dimethyl thiazole-2-yl(-2,5-diphenyl tetrazolium bromide)] (MTT) dissolved in PBS as MTT reagent were used.

HeLa cells

HeLa cells being used were from the collection of Cancer Chemoprevention Research Center (CCRC), Universitas Gadjah Mada. The cell line was a gift from Prof. Tatsuo Takeya, *Nara Institute of Science and Technology* (NAIST), Japan.

Cytotoxicity and combinatorial assay

MTT cytotoxicity assay was used to examine the effect of LEE alone and in combination with doxorubicin on HeLa cells.

5x10³ HeLa cells/well was distributed into 96-well plate (Iwaki) and incubated in 37°C with 5% CO₂ (Heraeus) for 24 hours. In combinatorial assay, concentration of 5, 50, and 250 µg/ml for LEE and 100, 250 nM for doxorubicin were used. After 24 hours incubation, MTT reagent was applied, followed by 4 hours incubation. 10% SDS in 0.1N HCl as stopper reagent was then applied. Plate was then kept with protection from light overnight, continued with absorbance determination (λ 595 nm) using ELISA reader (Bio-Rad).

Analysis

The cells' viability was calculated based on the formula as follow:

$$\% \text{ viability} = \frac{\text{absorbance of treated cells} - \text{absorbance of control media}}{\text{absorbance of control cells} - \text{absorbance of control media}} \times 100\%$$

The cells' viability of LEE combined with doxorubicin treated cells then compared to LEE treated alone.

RESULTS

Single and combinatorial treatment of LEE and doxorubicin on HeLa cells gave the viability shown in Table 1, and being plotted in diagram (Figure 1). It was shown that four concentration of combinations shows synergism on HeLa cells, represented with less cells' viability of combinatorial treatment compared to single treatment.

Cells' morphology after treatment was also observed (Figure 2). Treatment of LEE or doxorubicin alone led to cells' morphological change pointed by white arrows, respectively (Figure 2(b) and 2(c)). Combination of them caused more changes compared to single treated cells (Figure 2(d)). While control cells showed no changing in cells' morphology representing cells' death (Figure 2(a)). Hence, synergism of combinatorial treatment was observed.

Table 1. Cell viability (%) of single and combinatorial treatment of LEE and doxorubicin on HeLa cells

LEE concentration (µg/ml)	Doxorubicin (nM)		
	0	100	250
0	100	102.49	99.33
5	110.47	107.98	92.68
50	99.50	85.62	86.28
250	64.51	65.84	46.97

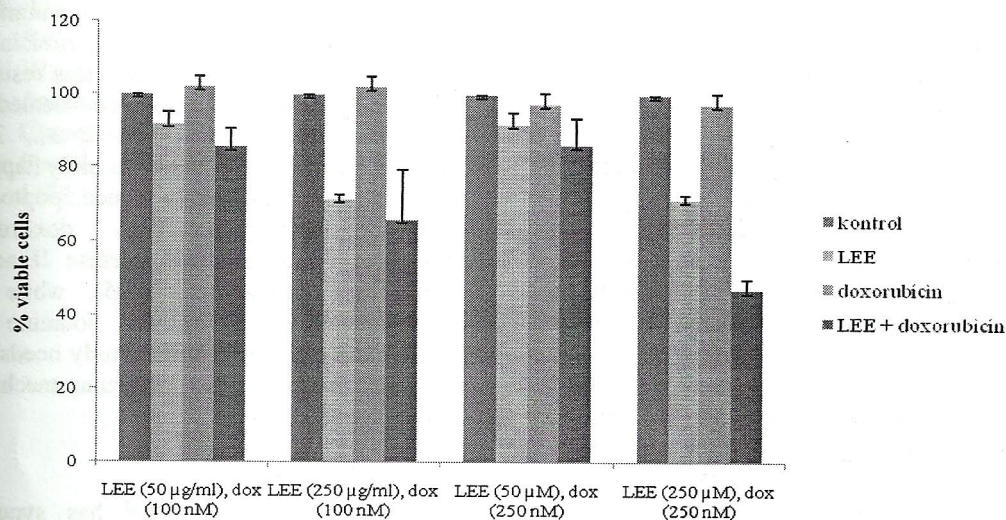


Figure 1. Combinational effect of LEE and doxorubicin. Test were carried out by incubating 5×10^3 MCF-7 cells/well with concentration 50 µg LEE/ml, 250 µg LEE/ml, and doxorubicin 100 nM, 250 nM for 24 hours. After 24 hour, cell were added by MTT reagent to obtain the absorbance representing cell viability. Combination treatment of LEE 250 µg/ml and doxorubicin 250 nM give optimal reduction of cell viability.

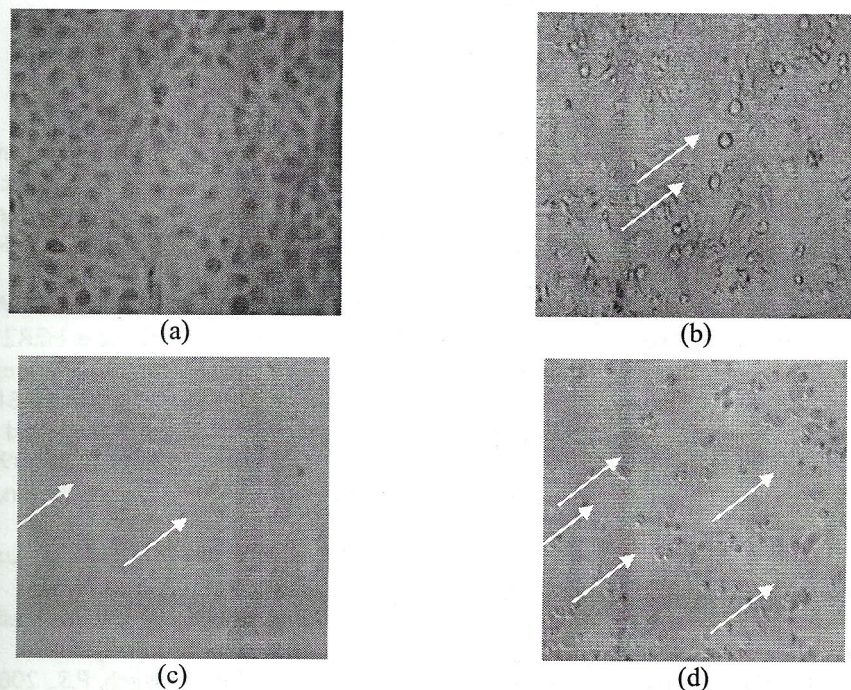


Figure 2. Combination of doxorubicin and LEE showed synergism on HeLa cells; (a) cell control; 24-hours treatment of (b) 250 µg/ml LEE; (c) 250nM doxorubicin; (d) 250nM doxorubicin in combination with 250 µg/ml LEE. Treatment of LEE and doxorubicin alone brought cells to death, shown by change in cells' morphology pointed by white arrows, respectively (b and c). Combination of them caused more cells' death compared to single treated cells (d), showing synergism. While control cells showed no change in cells' morphology (a). Observation was done by using inverted microscope at 300x magnification.

DISCUSSION

Leunca ethanolic extract alone is cytotoxic against HeLa cells with IC_{50} 227 µg/mL

(Istiaji *et al.*, 2010). When it is combined with doxorubicin, it performed synergistic effect. The cells' viability is decreasing compared to LEE treated cells, meaning that the combination gave

inhibition of HeLa cells' growth. LEE may contain solanine and solamargine (Everist, 1974; Weller and Phipps, 1979). Solanine and β -2-solamargine suppress Bcl-2 expression (Ji *et al.*, 2008; Liu *et al.*, 2004). Bcl-2 is an antiapoptotic protein. Overexpression of Bcl-2 prevented the efflux of cytochrome c from the mitochondria and the initiation of apoptosis (Yang *et al.*, 1997). When it suppressed, cells will undergo apoptosis easier. Cytosolic cytochrome c will be larger when Bcl-2 is down regulated. This will lead to initiation of apoptosis (Yang *et al.*, 1997). Liang *et al.* (2008) also proved that β -2-solamargine induced apoptosis through mitochondrial pathway, rendering the correlation between Bcl-2 suppression and apoptosis induction via classical pathway. The TNFRs induction on lung cancer cells (Liu *et al.*, 2004) probably could be other mechanism on HeLa cells' growth inhibition. The molecular targeted protein of LEE should be determined briefly.

The combination of LEE and doxorubicin exhibited synergism on inhibition of

HeLa cells' growth. To have the same number of cells' viability, we'll need less doxorubicin. This will lessenside effects as well as less resistance phenomenon found in cancer cells treated with chemotherapeutic agent (Kufe *et al.*, 2003). Synergism of the combination possibly happened due to the ability of LEE to induce apoptosis by different mechanism with doxorubicin. Doxorubicin inhibits topoisomerase II activity (Potter and Rabinovitch, 2005), while LEE induces apoptosis via its solanine and solamargine activities. Further study needs to be conducted to determined molecular mechanism underlying this activity.

CONCLUSION

We can conclude that LEE has synergism activity when combined with doxorubicin on HeLa cells.

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