E-ISSN: 2455-3891 P-ISSN: 0974-2441

Digital Repository Universitas Jember

Vol 12, issue 1, 2019

An Open Access Peer Reviewed Journal

Asian Journal of Pharmaceutical & Clinical Research

Citation Abbreviation: Asian J. Pharm. Clin. Res. www.innovareacademics.in



Scanned by CamScanner

Asian Journal of Pharmaceutical and Clinical Research

ISSN- 0974-2441

Editorial Board

Editor-in-Chief

Dr. Anurekha Jain

Dept. of Pharmaceutical Sciences, Jyoti Mahila Vidyapeeth University, Jaipur, Rajasthan Email: editor@ajpcr.com

Associate Editor

Dr. Neeraj Upmanyu

Peoples Institute of Pharmacy & Research Center, Bhopal, MP, India

Dr. Vikas Sharma

Shri Rawatpura Sarkar Institute of Pharmacy, Datiya, MP, India

Assistant Editor

Dr. Vimal Kumar Jain

Institute of Pharmacy, Nirmal University, Ahemdabad, Gujarat, India

Dr. Rupesh Kumar Gautam *

ADINA Institute of Pharmaceutical Sciences, Sagar, MP, India

Editorial Board Members

Prof Mohamed Eddouks, Morocco Prof. Ahmed Osman, Malaysia Prof. B. M. Patil, India Prof. Syed Azhar Syed Sulaiman, Malaysia Prof. Joao B. Calixto, Brazil Prof Hakan Arslan, Turkey Dr. Rajesh Kumar, India Dr. Selvakumar Dakshnamurthy, Japan

- Dr. Ritu Mehra Gilhotra, India Dr. Supriya Shrihari Shidhaye India Dr. Harish Dureja, India Dr. Molly Mathew, India Dr. K. Mahour, India Dr. Vijay Dhondiram Wagh, India Dr. Subrat Kar, India Dr. Vikas Anand Saharan, India Dr. Debasish Maiti, India Dr. Amer A. Taqa, India
- Dr. Rashad Musleh Alamer, Morocco Dr. Shubhamoy Ghosh, India Dr. Vikas Anand Saharan, India Mr. Mohammad Nurul Amin, India Mr. Mohd Abdul Hadi, India Mrs. Manjula Sunkara, USA Mr. Ravikiran Donthu, UIUC Mr. Neeraj Gilhotra, India Mr. Abhay R. Shirode India Mr. Manish Kumar, India

Addresss

Published By

Innovare Academic Sciences Pvt Ltd B-11 Housing Colony In Front Of Bima Hospital,Nai Abadi,Mandsaur M.P. 458001 Email: editor@ajpcr.com

Print By

Fun & Art

29-Nagar Palika Complex, Gandhi chouraha, Mandsaur M.P. 458001

Branch Office

Innovare Academic Sciences Pvt Ltd T-8, Mahaveer Apartment, Near SIRT College, Ayodhya Bypaas, Bhopal-462041, MP, India URL: http://innovareacademics.in | Email: iajournals@gmail.com | Contact: +91 810 972 5561

AJPCR Scope

AJPCR (Asian J Pharm Clin Res) is peer reviewed, Bimonthly (Onward August 2014) open access Journal. This journal publishes original research in the field of Pharmaceutical and Clinical Sciences. The Journal has been designed to cover all the fields of research, which has any correlation and impact on Pharmaceutical Science. It aims to publish all the original research in field of science so a correlation can be made between these researches. Knowledge gained by such researches can be exposed to all and it can be brought in real utilization as all the branches of science are correlated and will assist all the researchers to potentiate their research capabilities.

Vol 12. Issue 1, 2019

Abstracting and Indexing

Abstracting and Indexing- Google Scholar, Scopus, Elsevier, EBSCO, EMBASE, SCI mago (SJR), CAS, CASSI (American Chemical Society), DOAJ (Directory of Open Acess Journal), Index Copernicus, ICAAP (International Consortium for the Advancement of Academic Publication), Open-J-Gate, Socolar. ASIAN JOURNAL OF PHARMACEUTICAL AND CLINICAL RESEARCH



VASOPROTECTIVE EFFECT OF *PHYSALIS ANGULATA* L. LEAF WATER EXTRACT ON KIDNEY OF NΩ-NITRO-L-ARGININE METHYL ESTER-INDUCED ENDOTHELIAL DYSFUNCTION RAT MODEL

ZAHRAH FEBIANTI^{1,2}, NUR PERMATASARI^{3*}, SETYAWATI SOEHARTO³

¹Department of Biomedical Sciences, Faculty of Medicine, Brawijaya University, Malang 65145, Indonesia. ²Department of Biochemistry, Faculty of Medicine, University of Jember, 68121, Indonesia. ³Department of Pharmacology, Faculty of Medicine, Brawijaya University, Malang 65145, Indonesia. Email: nungky.permatasari@gmail.com

Received: 20 September 2018, Revised and Accepted: 26 November 2018

ABSTRACT

Objective: This study was to investigate the effect of *Physalis angulata* L. leaf water extract on vascular rarefaction, oxidative stress, and inflammation on the kidney of Nω-nitro-L-arginine methyl ester (L-NAME)-induced male Wistar rats.

Methods: A total of 25 male Wistar rats were divided into five equal groups (normal control: 40 mg/kg/day of normal saline; L-NAME group; and treatments I, II, and III: L-NAME plus *P. angulata* L. leaf water extract doses 500 mg/kg/day, 1500 mg/kg/day, and 2500 mg/kg/day, respectively). Endothelial dysfunction was induced by 40 mg/kg/day L-NAME intraperitoneally. The treatment lasts for 15 days. Vascular rarefaction was indicated by the decrease in vascular density, which considered as vascular number per high-power field in hematoxylin-eosin staining preparation. Kidney oxidative stress test was performed by measuring malondialdehyde (MDA) level with thiobarbituric acid reactive substances assay. The inflammatory marker was the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) expression which examined using an immunohistochemical method with an antibody against p65.

Results: At the dose of 500 mg/kg/day and 1500 mg/kg/day, *P* angulata leaf water extract supplementation increased the vascular density, decreased the MDA level, and decreased the NF-KB expression compared to the L-NAME group.

Conclusion: The administration of *P. angulata* L. leaf water extract in particular concentration has a vasoprotective effect by preventing kidney vascular rarefaction, oxidative stress, and inflammation on L-NAME-induced male Wistar rat.

Keywords: Endothelial dysfunction, Kidney, Malondialdehyde, Nuclear factor kappa-light-chain-enhancer of activated B cells, Nω-nitro-L-arginine methyl ester, *Physalis angulata* leaf, Vasoprotective.

© 2019 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (http://creativecommons. org/licenses/by/4. 0/) DOI: http://dx.doi.org/10.22159/ajpcr.2019.v12i1.29857

INTRODUCTION

Endothelial dysfunction is indicated by the decreased bioavailability of vasodilators, primarily nitric oxide (NO) [1,2]. In the kidney, NO has various important functions, such as regulation of renal hemodynamics, regulation of glomerular microcirculation and salt balance, blunting of tubuloglomerular feedback, and modulation of renal sympathetic nerve activity [3,4]. The inhibition of NO synthesis by N ω -nitro-L-arginine methyl ester (L-NAME) in rats leads to endothelial vasoconstriction and activation indicated by pro-inflammatory, proliferative, and procoagulant conditions [4,5]. It results in severe hypertension and causes kidney damage [3,4].

Endothelial dysfunction creates an imbalance between NO and reactive oxygen species (ROS) which leads to oxidative stress [6]. ROS can attack various biomolecular components of the cells, such as lipid, which causes lipid peroxidation (LPO). This reaction released LPO products, such as malondialdehyde (MDA) [3]. ROS also activates one of the transcription factors, such as nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B), which will subsequently induce the synthesis of inflammatory mediators, and vice versa [7]. Excessive NF- κ B activation leads to excessive inflammation and cell damage [8]. The damaged kidney vascular cells lead to structural changes called renal vascular rarefaction indicated by the decrease in vascular density [9,10]. Kidney capillary rarefaction is associated with aging and renal fibrosis and is an indicator of impaired renal function [11]. Therefore, preventing kidney vascular rarefaction, oxidative stress, and inflammation is important in the management of endothelial dysfunction associated kidney diseases.

Ciplukan (*Physalis angulata* L.) leaf is known to have high antioxidant effects *in vitro* [12]. The previous study proved that

P. angulata leaf extract contains *Physalin*, a class of secosteroids. This compound could increase the release of NO from endothelial cell *in vitro*. This increase of NO release is thought to occur through genomic effects by increasing the expression of endothelial NO synthase (eNOS) and inducible NO synthase and non-genomic effects by increasing cytosolic calcium [13]. Another study proved the target of *P. angulata* leaf water extract in NO synthesis pathway through increased levels of vascular endothelial growth factor (VEGF) and eNOS [14].

The objective of this study is to investigate the effects of *P. angulata* L. leaf water extract, which contains physalin, withanolides [13,15,16], and flavonoid [17] on vascular rarefaction, oxidative stress, and inflammation in the kidney of NOS-inhibited Wistar rats by L-NAME.

METHODS

Animal preparation

A total of 25 male Wistar rats, weighing 250-300 g, were placed in a quiet room with cage temperature $21\pm2^{\circ}$ C, in which a 12-12 h light-dark cycle was maintained. They were fed and watered by *ad libitum*. After 7 days of acclimatization, Wistar rats were divided into five groups (control negative: normal saline 0.9%, i.p; L-NAME group: 40 mg/kg/day L-NAME i.p; and L-NAME 40 mg/kg/day, i.p. + *P* angulata leaf water extract doses of 500, 1500, and 2500 mg/kg/day by gavage, respectively). The treatment lasts for 15 days [3,18,19]. The dose of water extract of *P* angulata leaves was determined based on the previous study [20]. The procedures were approved by the Health Research Ethics Committee of Brawijaya University (No.114/ EC/KEPK/03/2017).

Extract preparation

P. angulata leaves were obtained from Balai Penelitian Tanaman Obat dan Rempah, Lembang, Indonesia. The sample of the herbs was determined and confirmed as *P. angulata* species by the Laboratory of School of Life Sciences and Technology, Bandung Institute of Technology, Indonesia. As much as 10 g of *P. angulata* leaves dry powder were soaked in 100 ml of boiled water for an hour. The solution was filtered from the precipitated product by cotton cloth to obtain 80 ml thick extract. The same process was repeated for the remaining product. It was soaked again in 30 ml boiled water for an hour and then filtered to obtain 20 ml extract. From this procedure, we got 100 ml *P. angulata* leaf extract with a concentration of 10% (w/v).

Determination of MDA level

Kidney LPO was used as an indicator of oxidative stress [21]. It was measured according to the concentration of thiobarbituric acid (TBA) reactive substances. The amount of produced MDA was used as an index of LPO [22]. MDA and TBA react and produce pink pigment with a maximum absorption at 535 nm [23]. Tissue homogenate (0.1 ml) was mixed with 2 ml reagent consisting of 0.37% TBA, 0.25 N HCl, and 15% trichloroacetic acid with 1:1:1 ratio. The mixture was then placed in a boiling water bath for 15 min. After cooling, the mixture was centrifuged at room temperature for 10 min and the absorbance of the clear supernatant was read at 535 nm [24]. Data were expressed as nm/300 mg tissue.

Immunohistochemical preparation and evaluation

At the end of the experimental period, the rat kidneys were removed, fixed in 10% buffered formalin solution, and then embedded in paraffin. Each kidney was cut in a sagittal section into two halves. Paraffin kidney sections (5 mm) were prepared for immunohistochemical examination, and another was stained with hematoxylin and eosin. For immunohistochemical analysis of NF- κ B, antigen unmasking was performed by heating the sections in citrate buffer, pH 6.0, using a water bath 95°C for 20 min. Sections were incubated overnight with primary antibodies (1:100 polyclonal p65, Santa Cruz Biotechnology) at 4°C. The detection of immunopositive cells used the avidin-biotin-peroxidase complex method. Immunoreactivity was visualized with diaminobenzidine. Hematoxylin was used as the counterstain. Negative controls consisted of each case in which the primary antibody was omitted.

Renal sections were scored for the presence of p65 in renal cells, whether in the cytoplasm or in the nucleus. Renal cells were quantitatively measured by counting at least 20 randomly selected high-power fields (×400) in the cortex and outer medulla area. The final score obtained was expressed as the number of positive cells per high-power fields [25].

Histopathological evaluation

The vascular density of the kidneys was evaluated by counting the vascular in 20 randomly selected field sites in the renal cortex and outer medulla with a magnification of ×400. It will be considered as vascular if there is a lumen containing erythrocytes and coated by endothelial cell(s) [26]. Data were expressed as the number of vascular per high-power field.

Statistical analysis

The data were analyzed with SPSS 23.0 for Windows using independent t-test to compare the control and L-NAME group. One-way analysis of variance was used to compare the L-NAME group and the treatment groups. Differences between group were determined using *post hoc* analysis in which the significance level was described as p<0.05.

RESULTS

The L-NAME effect on vascular density, MDA levels, and NF- $\!\kappa B$ expression in rat kidney

The effects of L-NAME on vascular density, MDA levels, and NF- κ B expression in rat kidney are shown in Fig. 1a-c, respectively. L-NAME

tended to decrease the vascular density in rat kidney compared to the control group, but it was not statistically significant, while the MDA level and NF- κ B expression tended to increase in the L-NAME group compared to the control group, but it was not significantly different.

The effect of *P. angulata* leaf water extract on kidney vascular density

Administration of *P. angulata* leaf water extract at dose 1 (500 mg/kg/day) and dose 2 (1500 mg/kg/day) could increase vascular number significantly to L-NAME group. Increasing the dose of *P. angulata* leaf water extract tended to decrease the vascular number in renal cortex compared to dose 1 (500 mg/kg/day). At dose 3 (2500 mg/kg/day), there was significant decrease on vascular number compared to dose 1 (500 mg/kg/day) but not significant to dose 2 (1500 mg/kg/day) (Figs. 2 and 3).

The effect of *P. angulata* leaf water extract on kidney MDA level

MDA level is an important indicator of oxidant status. The data of MDA level in L-NAME plus *P. angulata* leaf water extract with various dose groups are indicated in Fig. 4. There was a decrease in MDA level of *P. angulata* leaf water extract at dose 1 of 500 mg/kg/day compared to L-NAME group. Increasing the dose of the *P. angulata* leaf extract at dose 2 (1500 mg/kg/day) and 2500 mg/kg/day tended to increase the MDA level compared to the lowest dose (Fig. 4).

The effect of *P. angulata* leaf water extract on NF-κB p65 expression

The effect of L-NAME and *P. angulata* leaf water extract on NF-kB expression is shown in Fig. 5. It was shown that the administration of 500 mg/kg/day (dose 1) *P. angulata* leaf water extract tends to decrease p65 expression on rat kidney, but it was not significant. Administration of *P. angulata* leaf water extract at higher doses (doses 2 and 3) tends to increase p65 NF- κ B expression compared to dose 1 (Figs. 5 and 6).

DISCUSSION

This study showed that the vascular density in the renal cortex was decreased after 15 days of L-NAME induction followed by the increasing level of MDA and NF-kB expression (Fig. 1). It was in accordance with the previous studies, showing that L-NAME may cause endothelial dysfunction by lowering NO which promotes thrombosis, vasospasm, vascular inflammation, and proliferation of vascular smooth muscle cells [1]. Oxidative stress also contributes to the mechanisms of endothelial dysfunction. L-NAME may increase oxidative stress by enhancing nicotinamide adenine dinucleotide phosphate oxidase expression [27,28]. Cell membranes composed of poly-unsaturated fatty acids are particularly susceptible to oxidative attack, which resulted in the changes of permeability, membrane fluidity, and cellular metabolic functions. MDA, one of the LPO products, was found to increase in oxidative stress state [21]. L-NAME can induce inflammation and kidney damage by increasing angiotensin (AT) 2 stimulation to AT1-receptor [29] so that it activates the transcription factor NF-KB [30]. These processes finally will lead to microvascular rarefaction [9,31], which was indicated by a decreased in vascular density [10].

The insignificant result of all three variables in the L-NAME group compared to the control group can be caused by a relatively short duration of treatment and/or less L-NAME dose being used. A study by Cipolla *et al.* [32] showed that it took 5 weeks of L-NAME administration at a dose of 0.5 g/L drinking water to cause loss of vascular (rarefaction) structures in the Sprague-Dawley rat brain capillaries. Meanwhile, the other studies were performed for 7–8 weeks to make significant renal damage and inflammation [4,33-36]. The other needed 28 days to significantly increase rat kidney MDA level [37-39]. These studies indicate that the kidney needs a longer duration than 15 days of L-NAME administration to significantly decrease the vascular density and increase the MDA level as well as the NF- κ B expression. Kidney endurance may contribute to this phenomenon. The kidney can protect itself from L-NAME-induced hypertension by its autoregulation mechanism. This autoregulation mechanism causes kidney blood

Digital Repository Universitas Jember Asian J Pharm Clin Res, Vol 12, Issue 1, 2019, 432-437

14 0.7 40 (jd 35 12 0.6 (cells/ MDA level (ng/300 mg) 70 0.7 70 70 70 70 (id10 Vascular density နို 25 vascular number 8 5 NF-KB positive 0 6 01 2 p65 5 n n Control (-) Control (-) Control L-NAME

Fig. 1: Mean of vascular density (a), mean of malondialdehyde level (b), and mean of p65 expression (c) control (without Nω-nitro-L-arginine methyl ester [L-NAME]) and L-NAME (40 mg/kg/day). Data were presented as mean ± standard error of the mean and analyzed using independent t-test



Fig. 2: Mean of kidney cortex vascular density. *p<0.05 compared to the Nω-nitro-L-arginine methyl ester (L-NAME) group. #p<0.05 compared to L-NAME + *Physalis angulata* dose 3. Data were analyzed using analysis of variance and *post hoc* test



Fig. 3: Kidney cortex of negative control group (a), Nω-nitro-Larginine methyl ester (L-NAME) group (b), L-NAME+extract dose 1 group (c), L-NAME + extract dose 2 group (d), L-NAME + extract dose 3 group (e). The red arrow indicates vascular. HE, ×400, inset ×1000, scale bar: 20 μm

pressure to be maintained normally despite an increase in systemic blood pressure [40].

Moreover, kidney cells can protect themselves from oxidative stress by synthesizing antioxidant enzymes. A study mentioned that renal ischemia would increase renal antioxidant enzymes [41]. This study



Fig. 4: Mean of kidney malondialdehyde level. Data were analyzed using the analysis of variance test and *post hoc* test







Fig. 6: Kidney p65 nuclear factor kappa light chain enhancer of activated B cells expression of the negative control group (a),
Nω-nitro-L-arginine methyl ester (L-NAME) group (b), and L-NAME + extract dose 1 group (c), L-NAME + extract dose 2 group (d),
L-NAME + extract dose 3 group (e). IHC, ×400, scale bar: 20 μm

showed that MDA levels in the L-NAME group did not increase significantly at the end of the treatment (Fig. 1b). It indicates that the antioxidant enzymes can still compensate for the free radical generated by L-NAME induction. Furthermore, AT2 also plays a role in kidney endurance [42]. The balance of AT2 effects on AT1 and AT2 receptors has an important impact on inducing kidney injury due to L-NAME administration. AT2 stimulation to AT1-receptor induces pro-oxidant and pro-inflammatory effects [43]. Meanwhile, AT2 stimulation to AT2-receptor counteracts those effects [44]. AT2-receptor stimulation can increase NO levels [45], possibly through direct stimulation of NOS

Asian J Pharm Clin Res, Vol 12, Issue 1, 2019, 432-437

and bradykinin pathway [46]. The AT2-receptor stimulation reduces inflammatory response through JAK/STAT inhibition, NF- κ B inhibition, and COX2 synthesis inhibition [47]. Therefore, in this study, we assumed that the reduction of NO levels induced by L-NAME still can be compensated by AT2 stimulation to AT2-receptor. Thus, the vascular density in the L-NAME group decreases insignificantly, following MDA level and NF- κ B expression which increase insignificantly when compared to the control group (Fig. 1a-c).

Ciplukan (P. angulata) was reported as an important herbal medicine in the Indian Traditional System of Medicine [15,16]. Qualitative analysis of the content of P. angulata leaf water extract and ethanol extract found the presence of flavonoids, saponins, terpenoids, polyphenols, tannins, alkaloids, and steroids [48,49]. Our result showed that the treatment groups receiving 500 mg/kg/day and 1500 mg/kg/day P. angulata leaf water extract had significantly higher vascular density compared to the L-NAME group (Figs. 2 and 3). Therefore, it can be assumed that supplementation of P. angulata leaf water extract can prevent L-NAME-induced vascular rarefaction. This result is in accordance with a previous study, showing that P. angulata could increase NO level of endothelial cell culture [13]. NO has a vasoprotective effect by preventing endothelial cell apoptosis [50] and by inhibiting caspase through S-nitrosylation of cysteine residues [51]. Adequate NO level promotes neovascularization, one of which through the VEGF pathway and fibroblast growth factor [52]. NO stimulates endothelial migration by inducing endothelial cell podokinesis, increasing the expression of $\alpha v\beta 3$, and enhancing dissolution of the extracellular matrix through the fibroblast growth factor-induced upregulation of urokinase-type plasminogen activator [53]. Furthermore, NO may suppress the production of angiostatin, an endogenous antagonist of angiogenesis [54]. Sulistyowati also proved that the effect of Ciplukan (P. angulata L.) water extract on NO synthesis pathway was by increasing VEGF levels [14]. VEGF was known as a substance needs to promote vascularization [52,55].

P. angulata leaf water extract has a high antioxidant effect *in vitro* [12, 16]. This might be due to the flavonoid content of *P. angulata* which are 5-Methoxy-6,7-methylenedioxyflavone and 5,6,7-trimethoxyflavone [56]. Flavonoids prevent tissue damage from free radicals through various mechanisms. First, it reacts with free radical molecules directly because the flavonoids have hydroxyl groups with high reactivity. Second, it inhibits xanthine oxidase to prevent superoxide formation when reoxygenation occurs [57]. In accordance with those previous studies, the addition of 500 mg/kg/day and 1500 mg/kg/day *P. angulata* leaf water extract was able to lower the MDA level of L-NAME-induced Wistar rat (Fig. 4). Therefore, it can be assumed that supplementation of *P. angulata* leaf water extract can prevent oxidative stress in the kidney of endothelial dysfunction Wistar rat model.

Treatment with P. angulata leaf water extract at a dose of 500 mg/kg/day and 1500 mg/kg/day also decreased p65 NF-κB expression (Figs. 5 and 6). The previous study reported that P. angulata leaves have anti-inflammatory effects through tumor necrosis factor- α (TNF- α) inhibition [16,20]. It was known that TNF- α can trigger the classic pathway of NF-KB activation, resulting in an inflammatory response. This is reinforced by the research of Grumbach et al. [58], stating that NO can inhibit NF-KB activation in vitro. NF-KB activation will be manifested as a total increase in the expression of p65 proteins of the NF-kB complex [59]. The interesting result from the immunohistochemistry analysis above is that the p65 expression is never found in glomerular cells (Fig. 6). It indicates the involvement of glomerular autoregulation mechanism which protects the glomerulus from L-NAME-induced inflammation. Despite the changes in renal perfusion pressure, the autoregulatory mechanism keeps the renal blood flow and glomerular filtration rate constant [40].

Ciplukan (*P. angulata*) leaves have a unique characteristic. At lower dose (500 and 1500 mg/kg/day), it serves as proangiogenic,

antioxidant, and anti-inflammatory substance. Conversely, at high dose (2500 mg/kg/day), it showed to decrease vascular density (Fig. 2), increase MDA level (Fig. 4), and increase NF-kB expression (Fig. 5). It indicates that there was an excessive NO formation induced by highdose administration of *P. angulata* leaf water extract. This finding was in accordance with the previous study, showing that high NO concentration could promote endothelial cell damage [60,61]. NO cytotoxic effect is related to the chemical reactivity of peroxynitrite (ONOO-) formed from NO [62]. ONOO- caused persistent activation of NF-KB [63]. Conversely, the cytoprotective action of NO is attributed to the inhibition of NF-kB-mediated gene expression which produces ubiquitous anti-inflammatory activity [62]. The molecular mechanisms underlying the proapoptotic effect of high-dose NO remain speculative. The factors determining whether endothelial cells undergo apoptosis when exposed to NO include the amounts of NO, the different redox states of NO, and the local environment that may promote the further production of cytotoxic moieties such as ONOO-. The previous study showed that NO and ONOO- can damage DNA directly. The damaged DNA triggers the p53-dependent or p53-independent apoptotic cell death pathways which furthermore activates caspases [60]. The damaged cells are not able to synthesize VEGF again [31]. Finally, the decrease in VEGF led to vascular rarefaction [64].

Moreover, exogenous antioxidants can act as a double-edged sword, becoming antioxidant at low doses and prooxidant at high doses [65,66]. Therefore, increasing *P. angulata* leaf water extract dose will probably increase the antioxidant which reacts as a prooxidant. Another study by Nnamani *et al.* [67] reported that *P. angulata* leaves contain cyanide. Cyanide can cause oxidative stress in cells resulting in cell death [68,69]. Therefore, we hypothesized that a high concentration of *P. angulata* leaf water extract has a contrary effect, which is antiangiogenic, pro-oxidant, and pro-inflammatory. We also hypothesized that the high MDA level of the treatment group receiving 2500 mg/kg/day *P. angulata* leaf water extract might be caused by the effect of cyanide which becomes more dominant.

CONCLUSION

Based on those results, we concluded that the administration of *P. angulata* L. leaf water extract in particular concentrations has a vasoprotective effect by preventing kidney vascular rarefaction, oxidative stress, and inflammation on L-NAME-induced male Wistar rat. This study implies the importance of the optimal dose of *P. angulata* leaf water extract supplementation for the prevention of endothelial dysfunction-induced kidney injury. However, vascular rarefaction pathway is not only triggered by NF- κ B and oxidative stress but also triggered by several pathways such as VEGF and proapoptotic signaling for vascular rarefaction. To investigate this pathway, further studies are needed.

ACKNOWLEDGMENT

This research was supported by grants from The Ministry of Research, Technology, and Higher Education of the Republic of Indonesia through INSINAS 2016 funding program.

AUTHOR CONTRIBUTION

All the authors have the same contribution in this research (carried out the research, collected the data, analyzed the data, and formatted the manuscript).

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

REFERENCES

- Endemann DH, Schiffrin EL. Endothelial dysfunction. J Am Soc Nephrol 2004;15:1983-92.
- Förstermann U. Nitric oxide and oxidative stress in vascular disease. Pflugers Arch 2010;459:923-39.

Asian J Pharm Clin Res, Vol 12, Issue 1, 2019, 432-437

- Hadi HA, Carr CS, Al Suwaidi J. Endothelial dysfunction: Cardiovascular risk factors, therapy, and outcome. Vasc Health Risk Manag 2005;1:183-98.
- Higashi Y, Maruhashi T, Noma K, Kihara Y. Oxidative stress and endothelial dysfunction: Clinical evidence and therapeutic implications. Trends Cardiovasc Med 2014;24:165-9.
- Talas ZS, Ozdemir I, Ciftci O, Cakir O, Gulhan MF, Pasaoglu OM, et al. Role of propolis on biochemical parameters in kidney and heart tissues against L-NAME induced oxidative injury in rats. Clin Exp Hypertens 2014;36:492-6.
- Tsuchiya K, Tomita S, Ishizawa K, Abe S, Ikeda Y, Kihira Y, *et al.* Dietary nitrite ameliorates renal injury in L-NAME-induced hypertensive rats. Nitric Oxide 2010;22:98-103.
- Oeckinghaus A, Ghosh S. The NF-kappaB family of transcription factors and its regulation. Cold Spring Harb Perspect Biol 2009;1:a000034.
- Guzik TJ, Harrison DG. Endothelial NF-kappaB as a mediator of kidney damage: The missing link between systemic vascular and renal disease? Circ Res 2007;101:227-9.
- Goligorsky MS. Pathogenesis of endothelial cell dysfunction in chronic kidney disease: A retrospective and what the future may hold. Kidney Res Clin Pract 2015;34:76-82.
- Olufsen MS, Hill NA, Vaughan GD, Sainsbury C, Johnson M. Rarefaction and blood pressure in systemic and pulmonary arteries. J Fluid Mech 2012;705:280-305.
- Schmitt R, Melk A. Molecular mechanisms of renal aging. Kidney Int 2017;92:569-79.
- 12. Kusumaningtyas RW, Lailya N, Limandha P. Potential of Ciplukan (*Physalis angulata* L.) as source of functional ingredient. Procedia Chem 2015;14:367-72.
- Nurdiana PN, Karyono S. Efek non genomik dan genomik ekstrak daun ceplukan (*Physalis minima* L) pada kultur sel endotel manusia (HUVECs). J Ilmu Ilmu Hayati 2010;22:14-9.
- Sulistiyowati Y. Sasaran Aksi Ekstrak Air Herba Ciplukan (*Physalis angulata* L) Terstandar Fisalin pada Jalur Sintesis Nitric Oxide Tikus Sprague Dawley Diinduksi Nicotinamide Dan Streptozotocin. Disertasi, Universitas Gadjah Mada; 2015.
- 15. Chothani DL, Vaghasiya HU. A phyto-pharmacological overview on *Physalis minima* Linn. Indian J Nat Prod Resour 2012;3:477-82.
- Sharma N, Bano A, Dhaliwal HS, Sharma V. A pharmacological comprehensive review on 'rassbhary' *Physalis angulata* (L.). Int J Pharm Pharm Sci 1987;7:30-4.
- Susanti RF, Kurnia K, Vania A, Reynaldo IJ. Total phenol, flavonoid and antioxidant activity of *Physalis angulata* leaves extract by subcritical water extraction. Mod Appl Sci 2015;9:190-8.
- Talas ZS, Ozdemir I, Cifici O, Cakir O. Propolis attenuates oxidative injury in brain and lung of nitric oxide synthase inhibited rats. J Pharm Care 2013;1:45-50.
- Selamoglu ZS, Ozdemir I, Ciftei O, Gulhan MF, Savei A. Antioxidant effect of ethanolic extract of propolis in liver of L-NAME treated rats. Adv Clin Exp Med 2015;24:227-32.
- Lestari B, Permatasari N, Rohman MS. Methanolic extract of ceplukan leaf (*Physalis minima* L.) attenuates ventricular fibrosis through inhibition of TNF-α in ovariectomized rats. Adv Pharmacol Sci 2016;2016:2428052.
- Gaweł S, Wardas M, Niedworok E, Wardas P. Malondialdehyde (MDA) as a lipid peroxidation marker. Wiad Lek 2004;57:453-5.
- Ayala A, Munoz MF, Arguelles S. Lipid peroxidation: Production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. Oxid Med Cell Longev 2014;2014:31.
- Grotto D, Maria LS, Valentini J, Paniz C, Garcia GS, Rocha JB, *et al.* Importance of the lipid peroxidation biomarkers and methodological aspects for malondialdehyde quantification. Quim Nova 2009;32: 169-74.
- 24. Ganie SA, Haq E, Hamid A, Qurishi Y, Mahmood Z, Zargar BA, et al. Carbon tetrachloride induced kidney and lung tissue damages and antioxidant activities of the aqueous rhizome extract of *Podophyllum hexandrum*. BMC Complement Altern Med 2011;11:17.
- Spandou E, Tsouchnikas I, Karkavelas G, Dounousi E, Simeonidou C, Guiba-Tziampiri O, *et al.* Erythropoietin attenuates renal injury in experimental acute renal failure ischaemic/reperfusion model. Nephrol Dial Transplant 2006;21:330-6.
- Moreno PR, Purushothaman KR, Sirol M, Levy AP, Fuster V. Neovascularization in human atherosclerosis. Circulation 2006;113:2245-52.
- Zabrecky A, Pham H, Koon A, Rueter B, Barsotti R, Young L, et al. The role of NADPH oxidase isoform 1 (NOX1) in L-NAME-induced leukocyte-endothelial interactions in rat mesenteric postcapillary

venules (Abstract). FASEB J 2016;30 Suppl:723-2.

- Touyz RM, Chen X, Tabet F, Yao G, He G, Quinn MT, et al. Expression of a functionally active gp91phox-containing neutrophil-type NAD(P) H oxidase in smooth muscle cells from human resistance arteries: Regulation by angiotensin II. Circ Res 2002;90:1205-13.
- Figueroa-Guillén ES, Castro-Moreno P, Rivera-Jardón FF, Gallardo-Ortiz IA, Ibarra-Barajas M, Godínez-Hernández D, *et al.* Angiotensin II pressor response in the L-NAME-induced hypertensive pithed rat: Role of the AT1 receptor. Proc West Pharmacol Soc 2009;52:54-6.
- Crowley SD. Linking angiotensin II to nuclear factor-κ light chain enhancer of activated B cells-induced cardiovascular damage: Bad CARMAs. Hypertension 2014;64:933-4.
- Chade AR. Renal vascular structure and rarefaction. Compr Physiol 2013;3:817-31.
- Cipolla MJ, Bishop N, Vinke RS, Godfrey JA. PPAR{gamma} activation prevents hypertensive remodeling of cerebral arteries and improves vascular function in female rats. Stroke 2010;41:1266-70.
- Pechanova O, Barta A, Vrankova S. The role of nuclear factor kappa B in L-NAME-induced hypertension. J Hypertens 2010;28:491-2.
- Peeri M, Habibian M, Azarbayjani MA, Hedayati M. Protective effect of aerobic exercise against L-NAME-induced kidney damage in rats. Arh Hig Rada Toksikol 2013;64:43-9.
- Vrankova S, Parohova J, Barta A, Janega P, Simko F, Pechanova O, et al. Effect of nuclear factor kappa B inhibition on L-NAME-induced hypertension and cardiovascular remodelling. J Hypertens 2010;28 Suppl 1:S45-9.
- 36. Gonzalez W, Fontaine V, Pueyo ME, Laquay N, Messika-Zeitoun D, Philippe M, et al. Molecular plasticity of vascular wall during N(G)nitro-L-arginine methyl ester-induced hypertension: Modulation of proinflammatory signals. Hypertension 2000;36:103-9.
- Abd-Eldayem AM, Farghaly HS, Abdel-Zaher AO. The nephroprotective effects of *Ginkgo biloba* extract (EGb761) against l-NG-nitroarginine methyl ester-induced hypertension in rats: Role of oxidative stress and inflammatory markers. J Curr Med Res Pract 2016;1:79-85.
- Valentine TM, Charles FN, Estella TA, Mauricette MA, Anatole AE, Julius OE, *et al.* Hydroethanolic extract of *Eribromao blongum* (*Malvaceae*) stem bark prevents hypertension, oxidative stress and dyslipidemia in L-NAME induced hypertension in wistar rats. J Dis Med Plants 2016;2:43.
- Khattab M, Ahmad M, Al-Shabanah OA, Raza M. Effects of losartan on blood pressure, oxidative stress, and nitrate/nitrite levels in the nitric oxide deficient hypertensive rats. Receptors Channels 2004;10:147-57.
- Carlström M, Wilcox CS, Arendshorst WJ. Renal autoregulation in health and disease. Physiol Rev 2015;95:405-511.
- Singh I, Gulati S, Orak JK, Singh AK. Expression of antioxidant enzymes in rat kidney during ischemia-reperfusion injury. Mol Cell Biochem 1993;125:97-104.
- Peluso AA, Santos RA, Unger T, Steckelings UM. The angiotensin Type 2 receptor and the kidney. Curr Opin Nephrol Hypertens 2017;26:36-42.
- 43. Sohn HY, Raff U, Hoffmann A, Gloe T, Heermeier K, Galle J, et al. Differential role of angiotensin II receptor subtypes on endothelial superoxide formation. Br J Pharmacol 2000;131:667-72.
- 44. Chabrashvili T, Kitiyakara C, Blau J, Karber A, Aslam S, Welch WJ, et al. Effects of ANG II Type 1 and 2 receptors on oxidative stress, renal NADPH oxidase, and SOD expression. Am J Physiol Regul Integr Comp Physiol 2003;285:R117-24.
- Matavelli LC, Huang J, Siragy HM. Angiotensin AT₂ receptor stimulation inhibits early renal inflammation in renovascular hypertension. Hypertension 2011;57:308-13.
- 46. Abadir PM, Foster DB, Crow M, Cooke CA, Rucker JJ, Jain A, et al. Identification and characterization of a functional mitochondrial angiotensin system. Proc Natl Acad Sci U S A 2011;108:14849-54.
- 47. Kaschina E, Namsolleck P, Unger T. AT2 receptors in cardiovascular and renal diseases. Pharmacol Res 2017;125:39-47.
- Karpagasundari C, Kulothungan S. Free radical scavenging activity of *Physalis minima* Linn. leaf extract (PMLE). J Med Plants Stud 2014;2:59-64.
- Patel T, Shah K, Jiwan K, Shrivastava N. Study on the antibacterial potential of *Physalis minima* Linn. Indian J Pharm Sci 2011;73:111-5.
- Yamashita T, Yamamoto E, Kataoka K, Nakamura T, Matsuba S, Tokutomi Y, *et al.* Apoptosis signal-regulating kinase-1 is involved in vascular endothelial and cardiac remodeling caused by nitric oxide deficiency. Hypertension 2007;50:519-24.
- Dimmeler S, Zeiher AM. Endothelial cell apoptosis in angiogenesis and vessel regression. Circ Res 2000;87:434-9.
- 52. Dulak J, Józkowicz A, Dembinska-Kiec A, Guevara I, Zdzienicka A, Zmudzinska-Grochot D, *et al.* Nitric oxide induces the synthesis of

Asian J Pharm Clin Res, Vol 12, Issue 1, 2019, 432-437

vascular endothelial growth factor by rat vascular smooth muscle cells. Arterioscler Thromb Vasc Biol 2000;20:659-66.

- Cooke JP, Losordo DW. Nitric oxide and angiogenesis. Circulation 2002;105:2133-5.
- Matsunaga T, Weihrauch DW, Moniz MC, Tessmer J, Warltier DC, Chilian WM, et al. Angiostatin inhibits coronary angiogenesis during impaired production of nitric oxide. Circulation 2002;105:2185-91.
- Nerkar D, Mukherjee A, Mehta BK, Banerjee S. Metabolic syndrome associated complications. Int J Pharm Sci 2015;7:22-5.
- Ser NA. Flavonoids from *Physalis minima*. Phytochemistry 1988;27:3708-9.
- Nijveldt RJ, van Nood E, van Hoorn DE, Boelens PG, van Norren K, van Leeuwen PA, *et al.* Flavonoids: A review of probable mechanisms of action and potential applications. Am J Clin Nutr 2001;74:418-25.
- Grumbach IM, Chen W, Mertens SA, Harrison DG. A negative feedback mechanism involving nitric oxide and nuclear factor kappa-B modulates endothelial nitric oxide synthase transcription. J Mol Cell Cardiol 2005;39:595-603.
- 59. Giani JF, Muñoz MC, Pons RA, Cao G, Toblli JE, Turyn D, et al. Angiotensin-(1-7) reduces proteinuria and diminishes structural damage in renal tissue of stroke-prone spontaneously hypertensive rats. Am J Physiol Renal Physiol 2011;300:F272-82.
- Shen YH, Wang XL, Wilcken DE. Nitric oxide induces and inhibits apoptosis through different pathways. FEBS Lett 1998;433:125-31.
- 61. Stefanec T. Endothelial apoptosis. Chest 2000;117:841-54.

- Hattori Y, Kasai K, Gross SS. NO suppresses while peroxynitrite sustains NF-kappaB: A paradigm to rationalize cytoprotective and cytotoxic actions attributed to NO. Cardiovasc Res 2004;63:31-40.
- Clancy RM, Gomez PF, Abramson SB. Nitric oxide sustains nuclear factor kappaB activation in cytokine-stimulated chondrocytes. Osteoarthritis Cartilage 2004;12:552-8.
- Lindenmeyer MT, Kretzler M, Boucherot A, Berra S, Yasuda Y, Henger A, *et al.* Interstitial vascular rarefaction and reduced VEGF-A expression in human diabetic nephropathy. J Am Soc Nephrol 2007;18:1765-76.
 Rahal A, Kumar A, Singh V, Yadav B, Tiwari R, Chakraborty S, *et al.*
- Rahal A, Kumar A, Singh V, Yadav B, Tiwari R, Chakraborty S, *et al.* Oxidative stress, prooxidants, and antioxidants: The interplay. Biomed Res Int 2014;2014:19.
- Bouayed J, Bohn T. Exogenous antioxidants-double-edged swords in cellular redox state: Health beneficial effects at physiologic doses versus deleterious effects at high doses. Oxid Med Cell Longev 2010;3:228-37.
- Nnamani C, Ani O, Belunwu G. Larvicidal effects of ethanol extracts on leaves and fruits of *Physalis angulata* L. on the larvae of anopheles mosquitoes from Ebonyi state, Nigeria, Anim Res Int 2010;6:1059-62.
- Shou Y, Gunasekar PG, Borowitz JL, Isom GE. Cyanide-induced apoptosis involves oxidative-stress-activated NF-kappaB in cortical neurons. Toxicol Appl Pharmacol 2000;164:196-205.
- Prabhakaran K, Li L, Borowitz JL, Isom GE. Cyanide induces different modes of death in cortical and mesencephalon cells. J Pharmacol Exp Ther 2002;303:510-9.

IAS Services

- Book Publication
- Journal Publication
- Journal Subscription
- Special Thematic Issue
- Copy-editing & Typesetting
- Conference Proceedings & Abstracts
- Conference/Seminar/Workshop/Symposia Event management

Why to publish in IAS?

- Rapid publication
- Reprints (on request)
- Unlimited space for your data
- Quality, reputation and high standard of peer review.
- Wide range of article publication
- Indexing in major bibliographic citation
- Provide high visibility due to open access

INNOVARE ACADEMICS SCIENCES JOURNALS

ository Universitas Jember

Asian Journal of Pharmaceutical & Clinical Research International Journal of Pharmacy & Pharmaceutical Sciences International Journal of Chemistry Research International journal of Applied Pharmaceutics International journal of Current Pharmaceutical Research Journal of Critical Reviews Innovare Journal of Medical Sciences Innovare Journal of Ayurvedic Sciences Innovare Journal of Health Sciences Innovare Journal of Engineering & Technology Innovare Journal of Social Sciences Innovare Journal of Business Management Innovare Journal of Education Innovare Journal of Agricultural Science Innovare Journal of Food Science Innovare Journal of Life Science Innovare Journal of Sciences

> Asian Journal of Pharmaceutical & Clinical Research B-11, In front of Beema Hospital, Nayi Abadi Mandsaur - 458001, MP, India Contact No: +91 940 661 2909 Email:editor@ajpcr.com www.innovareacademics.in

Innovare Academic Sciences Pvt. Ltd.

C All rights reserved

SINGLE USE

Scanned by CamScanner